OBSERVATIONS

Diabetes Screening Practices Among Individuals Aged 45 Years and Older

In 1997, the American Diabetes Association (ADA) adopted new recommendations for screening the general population aged ≥45 years for diabetes every 3 years with an emphasis on those at high risk for undiagnosed diabetes (1). Few studies have examined the extent to which this screening has been adopted. This report describes the results of a telephone survey of Montana residents aged ≥45 years to assess the diabetes screening practices in this population.

From October to December 1998, the Montana Department of Public Health and Human Services conducted a random household telephone survey of Montana residents aged ≥45 years living in 18 counties. Respondents indicated whether they had diabetes, the number of visits they made to a health care provider during the past year, their family history of diabetes, whether they had ever been told they had high cholesterol and/or high blood pressure, and their height and weight. Respondents were asked the following question to identify whether they had ever been screened for diabetes: “Glucose or sugar is a substance found in your blood. Have you ever had your blood glucose or sugar checked to see if you have diabetes?” When respondents answered “yes” to this question, they were asked to identify when screening was completed (“When was the last time you had your blood glucose or sugar level measured by a health care professional?”). The response categories for this question included within the past year, within the past 3 years, 3 years ago or having never been screened.

Respondents who reported being screened for diabetes during the past year were more likely to be age ≥65 years and to have a family history of diabetes, two or more visits to a health care provider during the past year, hypertension, and high cholesterol levels (Table 1). We found no association between recent screening and sex (40% men vs. 38% women), American Indian ancestry (41% yes vs. 38% no), or BMI (42% ≥27 kg/m² vs. 37% <27 kg/m²). Respondents with three or more risk factors (e.g., aged ≥45 years, American Indian ancestry, family history of diabetes, hypertension, high cholesterol, or BMI ≥27 kg/m²) were more likely to be screened for diabetes compared with respondents with only one risk factor (46 vs. 28%, respectively). However, 48% of individuals with two risk factors for diabetes and 37% of individuals with more than three risk factors had not been screened during the past 3 years.

Based on logistical regression analysis, three factors were associated with screening for diabetes during the past year: two or more visits to a health care provider during the past year (2.34 [95% CI 1.76-3.11]), high cholesterol level (1.37 [1.03-1.82]), and family history of diabetes (1.45 [1.12-1.89]). Respondents aged 45-54 years were less likely to report recent screening than those aged ≥65 years (0.62 [0.46-0.84]).

A limitation of this assessment is that these data are self-reported. Previous studies, however, have found that self-reports of conditions such as diabetes and hypertension are reliable (2,3). In addition, the survey was conducted by telephone and does not reflect the experience of individuals in Montana homes without telephones.

The findings suggest that diabetes screening is being adopted by physicians for individuals aged ≥45 years at risk for diabetes and that the ADA recommendations are being implemented in the general community. However, these data also indicate a need to develop strategies to encour-

Table 1—Characteristics of respondents aged ≥45 years reporting screening for diabetes in Montana in 1998

<table>
<thead>
<tr>
<th>Screening for diabetes</th>
<th>Past year</th>
<th>1-3 years</th>
<th>&gt;3 years or never</th>
</tr>
</thead>
<tbody>
<tr>
<td>n</td>
<td>430</td>
<td>160</td>
<td>520</td>
</tr>
<tr>
<td>Age (years)</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>45-54</td>
<td>155 (32)</td>
<td>80 (17)</td>
<td>251 (52)</td>
</tr>
<tr>
<td>55-64</td>
<td>95 (39)</td>
<td>41 (17)</td>
<td>108 (44)</td>
</tr>
<tr>
<td>≥65</td>
<td>180 (47)*</td>
<td>39 (10)</td>
<td>163 (43)</td>
</tr>
<tr>
<td>Family history of diabetes</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Yes</td>
<td>192 (45)*</td>
<td>74 (17)</td>
<td>162 (38)</td>
</tr>
<tr>
<td>No</td>
<td>235 (35)</td>
<td>85 (13)</td>
<td>352 (52)</td>
</tr>
<tr>
<td>Visits to a health care provider during the past year</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>≥2</td>
<td>317 (47)*</td>
<td>84 (12)</td>
<td>275 (41)</td>
</tr>
<tr>
<td>&lt;2</td>
<td>100 (24)</td>
<td>74 (18)</td>
<td>237 (38)</td>
</tr>
<tr>
<td>Hypertension</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Yes</td>
<td>149 (48)*</td>
<td>41 (13)</td>
<td>122 (39)</td>
</tr>
<tr>
<td>No</td>
<td>281 (35)</td>
<td>119 (15)</td>
<td>400 (50)</td>
</tr>
<tr>
<td>High cholesterol</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Yes</td>
<td>148 (47)*</td>
<td>43 (14)</td>
<td>123 (39)</td>
</tr>
<tr>
<td>No</td>
<td>280 (35)</td>
<td>116 (15)</td>
<td>396 (50)</td>
</tr>
</tbody>
</table>

Data are n (%). *P < 0.001.
HbA1c Is Not Recommended as a Screening Test for Diabetes in Cystic Fibrosis

In the June 1999 issue of Diabetes Care, Huckert et al. (1) recommend the use of HbA1c for early detection of cystic fibrosis-related diabetes (CFRD). Their recommendation is based on the finding that mean HbA1c is slightly higher in cystic fibrosis (CF) patients requiring insulin therapy, compared with CF patients with impaired or normal glucose tolerance. However, no data on the validity of this approach for the diagnosis of asymptomatic diabetes in patients with cystic fibrosis are presented.

Our group has a long-standing interest in the early diagnosis of CFRD, comparing fasting blood glucose levels with oral glucose tolerance test results (2). In our series, we have now 13 CF patients with newly diagnosed diabetes based on a 2-h venous plasma glucose value >200 mg/dl (11.1 mmol/l), and simultaneous determination of HbA1c (high-performance liquid chromatography method [Pharmacia, Erlangen, Germany], normal range 3.5–5.7%). Only 4 out of 13 CF patients (31%) diagnosed as diabetic according to the American Diabetes Association and World Health Organization criteria (3) had an HbA1c value above the normal range (individual values 5.9, 6.0, 6.3, and 6.6%). In nine diabetic CF patients with normal HbA1c values between 4.6 and 5.7% were encountered (mean ± SD, 5.1 ± 0.4%). The mean 2-h blood glucose value after ingestion of oral glucose was not significantly different between diabetic CF patients with normal HbA1c (263 ± 36 mg/dl, mean ± SD) and diabetic CF patients with elevated HbA1c (298 ± 34 mg/dl, Student’s t-test). These data clearly demonstrate that the determination of HbA1c, is not able to substitute for the oral glucose tolerance test in the early diagnosis of CFRD. Our findings are in agreement with several reports in the literature (4–6) as well as the 1998 consensus conference on CFRD (7). In addition to its low sensitivity when used as a diagnostic tool for the detection of CFRD, the measurement of HbA1c has the disadvantage of considerable interassay variability and lack of standardization. Therefore, we strongly advise against the use of glycosylated hemoglobin as a screening test for the early diagnosis of diabetes in patients with cystic fibrosis.

References

Pro115Gln Peroxisome Proliferator-Activated Receptor-γ and Obesity

Ristow et al. (1) reported an activating mutation in the peroxisome proliferator-activated receptor-γ gene (Pro115Gln PPAR-γ), which was present in 3% (4 of 121) of obese and 0% (0 of 238) of nonobese German Caucasians. These findings may have profound implications, particularly if the presence of this variant and its association with obesity are confirmed in other populations.

We performed polymerase chain reaction–restriction fragment length polymorphism analysis for the Pro115Gln PPAR-γ variant as described (1) on DNA samples from several independent populations, including lean and obese Caucasians from the Baltimore, Maryland, region; African-
Americans from Jackson, Mississippi, and Forsyth County, North Carolina; Pima Indians from Arizona; and Old Order Amish from Lancaster County, Pennsylvania (Table 1). A PCR fragment corresponding to gastric insulinotropic peptide, which has two known restriction sites for HindII, was mixed with each sample as a positive control. Among a total of 983 subjects (1,966 alleles), the Pro115Gln variant was not detected in a single subject.

These findings were unexpected because there is substantial overlap of gene pools of Caucasians from Central Europe and the Baltimore and Amish Caucasians studied (2). The German Caucasians studied by Ristow et al. were said to be unrelated and recruited from the Nordrhein-Westfalen region, but they may be a genetic isolate whose gene pool does not reflect that of other Caucasian populations. Alternatively, if the individuals carrying the mutation were related, the true frequency of the Pro115Gln PPAR-γ variant may have been overestimated. The absence or very low frequency of this variant has also been documented in Danish (3) and German (4) populations. This study is the first, to our knowledge, to examine American populations for this variant.

In summary, the study by Ristow et al. demonstrating that the Pro115Gln PPAR-γ variant is activating and can influence body weight in people is important. However, this variant appears to be absent or very rare in the American populations studied. Additional studies are required in other regions of Europe and the U.S. to further define the relevance of this interesting genetic variant to susceptibility to obesity.

**Table 1—Characteristics of subjects screened for Pro115Gln PPAR-γ**

<table>
<thead>
<tr>
<th>Population</th>
<th>n (alleles typed)</th>
<th>Age ± SD (years)</th>
<th>Female (%)</th>
<th>BMI ± SD (kg/m²)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Caucasians</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Baltimore Longitudinal Study on Aging</td>
<td>105 (210)</td>
<td>58.7 ± 15.3</td>
<td>53.3</td>
<td>24.8 ± 5.3</td>
</tr>
<tr>
<td>Johns Hopkins Weight Management Center</td>
<td>285 (570)</td>
<td>42.8 ± 12.0</td>
<td>65.3</td>
<td>38.8 ± 10.0</td>
</tr>
<tr>
<td>Amish, Lancaster, PA</td>
<td>173 (346)</td>
<td>45.7 ± 14.2</td>
<td>51.4</td>
<td>27.4 ± 8.9</td>
</tr>
<tr>
<td>African-Americans</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Atherosclerosis Risk in Communities Study</td>
<td>228 (456)</td>
<td>54.8 ± 5.9</td>
<td>60.0</td>
<td>31.4 ± 5.5</td>
</tr>
<tr>
<td>Pima Indians, Arizona</td>
<td>192 (384)</td>
<td>47.5 ± 14.8</td>
<td>—</td>
<td>32.6 ± 6.5</td>
</tr>
</tbody>
</table>

All non-Amish subjects were unrelated; Amish subjects were not first-degree relatives of each other.

**Acknowledgments**—This study was supported by NIH R01 DK-49692, the American Diabetes Association, GlaxoWellcome, the Baltimore Geriatrics Research and Education Clinical Center of the Baltimore Veterans Administration Medical Center.

**References**


**Insulin Secretion, Insulin Sensitivity, and Glucose Effectiveness in Nonobese Individuals With Varying Degrees of Glucose Tolerance**

Although it is well known that insulin secretion, insulin sensitivity, and glucose effectiveness are impaired in type 2 diabetic patients (1–5), little is known about the role of each of these factors individually on the evolution of type 2 diabetes. In this context, a major issue is that hyperglycemia per se impairs insulin secretion and insulin sensitivity and that obesity observed in type 2 diabetic patients per se causes insulin resistance (6,7). To overcome this problem, we studied 37 untreated nonobese subjects classified as having normal glucose tolerance (NGT) (n = 14; BMI 21.0 ± 0.5 kg/m² [range 18.1–23.6], mean ± SEM), impaired glucose tolerance (IGT) (n = 12; BMI 21.3 ± 0.8 kg/m² [17.0–26.9]), and type 2 diabetes (n = 11; fetal bovine serum 7.0 ± 0.5 mmol/l [range 5.2–9.9], BMI 20.0 ± 0.9 kg/m² [13.6–24.5]), based on the criteria of the World Health Organization (8). They were normotensive and had normal renal, hepatic, and thyroid function. Insulin sensitivity and glucose effectiveness were estimated by the minimal model approach (1–4). Insulin secretion was expressed as the area under the insulin curve between 0 and 10 min after an intravenous glucose injection (2). After the data were analyzed by one-way analysis of variance, Bonferroni correction was used to evaluate the differences between any two of the three groups we studied (9). No significant difference was observed in BMI among the three groups. Compared with the subjects with NGT, the subjects with
I GT had significantly lower insulin secretion (3.207 ± 557 vs. 1.895 ± 319 pmol · l⁻¹ · min⁻¹, \( P = 0.027 \)) and glucose effectiveness (0.023 ± 0.002 vs. 0.014 ± 0.002 min⁻¹, \( P < 0.001 \)). Insulin sensitivity index was lower in subjects with IGT (0.74 ± 0.13 min⁻¹ · pmol · l⁻¹) than in those with NGT (1.05 ± 0.14 min⁻¹ · pmol · l⁻¹), but was not statistically significant (\( P = 0.171 \)). In contrast, disposition index calculated by the product of insulin secretion and insulin sensitivity was significantly lower in subjects with IGT (1.197 ± 221) than in those with NGT (2.710 ± 371, \( P = 0.004 \)). On the other hand, patients with type 2 diabetes had significantly lower insulin secretion (212 ± 85 pmol · l⁻¹ · min⁻¹, \( P = 0.008 \)) compared with subjects with IGT. Although no significant difference was observed in insulin sensitivity index between subjects with type 2 diabetes and IGT (0.74 ± 0.13 vs. 1.09 ± 0.21 min⁻¹ · pmol · l⁻¹, \( P = 0.141 \)), disposition index was significantly diminished in type 2 diabetic patients as compared with subjects with IGT (224 ± 89 vs. 1.197 ± 221, \( P = 0.022 \)). Glucose effectiveness in type 2 diabetic patients (0.011 ± 0.001 min⁻¹) was similar to that in subjects with IGT (0.014 ± 0.002 min⁻¹, \( P = 0.307 \)) but was significantly lower than that in the subjects with NGT (\( P < 0.001 \)). From these results, the following may be hypothesized: 1) Impairments in insulin secretion and disposition index and decreased glucose effectiveness, but not insulin resistance, seem to constitute the basic characteristics of patients with IGT or type 2 diabetes in nonobese Japanese populations. 2) Risk factors worsening to type 2 diabetes in subjects with IGT are associated with further impairments in insulin secretion and disposition index, but not associated with further derangement in glucose effectiveness in Japanese populations.

References


Late-Onset Troglitazone-Induced Hepatic Dysfunction

Recently, Iwase et al. (1) reported a case of liver dysfunction occurring after 19 months of troglitazone therapy. Because it was thought before this report that the risk of liver dysfunction with troglitazone after 12 months was negligible, we wish to report another patient who took troglitazone intermittently and developed hepatic dysfunction after 18 months.

A 76-year-old white man with type 2 diabetes, ischemic heart disease (post angioplasty and stent placement), hypertension, degenerative joint disease, benign prostatic hypertrophy, dyslipidemia, and gastroesophageal reflux disease had troglitazone 400 mg daily added to his regimen of glimepiride 4 mg daily and metformin 500 mg b.i.d. because of poor glycemic control (HbA1c 8.9% [normal 4–6%]). The other medicines he used were aspirin and pravastatin. After 3 months of triple oral therapy, his HbA1c level dropped to 6.3%, and, after 6 months, to 6.0%. After 6 months, the patient’s HbA1c began to rise: 7.1% at 9 months, 7.9% at 1 year, and, 9.6% at 18 months.

Liver function tests were normal until 18 months, when his aspartate aminotransferase (AST) was found to be 64 (normal 0–37 U/l) and alanine aminotransferase (ALT) 68 (normal 7–56 U/l). The troglitazone regimen was discontinued, and testing for hepatitis B and C, hemachromatosis, autoimmune liver disease, and gallbladder disease were negative. Two months after discontinuing troglitazone, the patient’s AST and ALT had decreased to 50 and 64 U/l, and, after 3 months, had returned to normal at 33 and 36 U/l, respectively. His AST and ALT have remained normal since then and he has continued to take pravastatin, aspirin, metformin, and glimepiride.

When the patient was told to discontinue troglitazone, he admitted that he had been taking it only intermittently. He estimated that he took the drug regularly at first, but after the first 6 months, he took the drug only once or twice weekly on average. He gave the following three reasons for his lack of compliance: a lack of funds, a fear of liver disease, and a tendency to avoid taking drugs whenever possible.

This case, like the case described by Iwase et al., illustrates that the hepatic dysfunction caused by troglitazone can occur after 12 months and further supports the U.S. Food and Drug Administration’s current recommendation that quarterly liver function tests should be obtained when troglitazone utilization extends beyond 1 year (2).

In this case, could the onset of hepatic dysfunction have been delayed because the...
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D.S.H.B. and F.O. have served on an advisory panel for Sankyo Parke-Davis and have received consulting fees, research grant support, and honoraria for speaking engagements from Sankyo Parke-Davis.

References

Glyburide-Induced Hemolysis in Myelodysplastic Syndrome

Glyburide, also known as glibenclamide, is a widely used sulfonylurea to treat patients with type 2 diabetes. Hemolytic anemia is an extremely rare side effect of which there have been only a few reports (1–3). We describe a patient with myelodysplastic syndrome who presented with glyburide-induced hemolysis.

A 68-year-old man with a long history of type 2 diabetes presented with left foot cellulitis of 3 week’s duration. This patient was known to have slowly progressive panhypogonadism for 2 years, for which no work-ups had been performed. His medications included the following: glyburide, 5 mg per day; which he had taken for more than 1 year; buformine, 150 mg per day; and bglblose, 0.6 mg per day. He was afebrile, and the physical examination was normal, except for localized cellulitis on his left foot, for which he was started on intravenous antibiotics.

The laboratory studies revealed a white blood cell count of 3.3 × 10^9/l, hemoglobin 8.4 g/dl, platelet count 138 × 10^9/l, reticulocyte count 3.7%, mean corpuscle volume 96 fl, moderate anisocytosis, fasting plasma glucose 89 mg/dl, HbA1c 5.2%, lactate dehydrogenase 192 IU/l, total bilirubin 0.9 mg/dl, and haptoglobin <11 mg/dl. Red cell glucose-6-phosphate dehydrogenase level was adequate. Cold agglutinin test, Ham's test, and sugar water test were normal. Both direct and indirect Coombs' tests were negative. Red cell resistance to osmolarity was mildly low (Parpart's method). Urinalysis demonstrated no urobilinogen. Endoscopic studies did not reveal gastrointestinal bleeding. Ultrasonography of the abdomen showed no splenomegaly. The result of bone marrow aspiration was equivocal. On the basis of presumptive glyburide-induced hemolysis, glyburide was discontinued and the patient was switched to subcutaneous insulin on the seventh day. Thereafter, his hemoglobin level increased to 11.1 g/dl, reticulocyte count decreased to 1.3%, and anisocytosis disappeared promptly. He was discharged with insulin therapy after 1 month in the hospital, which is when the cellulitis resolved.

Three months later, his hemoglobin level was 10.0 g/dl and his haptoglobin remained low. Repeated bone marrow aspiration confirmed the diagnosis of myelodysplastic syndrome.

We conclude that this patient developed glyburide-induced hemolysis superimposed on red cell fragility secondary to an underlying bone marrow disorder. There have been several reports of hemolysis caused by sulfonylureas, most of which have been considered immune-mediated (1,2,4). Our case points to the possibility that glyburide could cause hemolysis by a non-immune-mediated mechanism. It is important to be aware of this potential side effect of glyburide in light of this medication’s widespread prescription, even though such a side effect is rare.

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Letters

Effects of Exposure at an Altitude of 3,000 m on Performance of Glucose Meters

Self-monitoring of blood glucose is mandatory for type 1 diabetic patients who participate in sports to adjust insulin dose and carbohydrate ingestion (1). Sports also include activities performed at moderately high altitudes, such as hiking or skiing. Capillary blood glucose monitors (BGMs) have been shown to underestimate blood glucose values at an altitude of 2,244 m (2) and at a simulated altitude of >2,000 m with temperature and humidity kept constant (3). The aim of the present study was to assess the accuracy of two BGMs at a moderately high altitude in which changes in temperature, humidity, and pH can result in errors in blood glucose determination (2).

Two BGMs, the LifeScan One Touch II (OT) (Ortho Diagnostics, Mipitas, CA) and the Glucometer Elite II (GE) (Bayer Diagnostics, Brussels, Belgium), were tested during a study on the effects of acute exposure at an altitude of 3,000 m and exercise on blood pressure and albumin excretion rate in six type 1 diabetic patients.

All subjects (four men and two women) were free of disease-related complications and in good and stable glycemic control (GHb 6.8 ± 1.1%). All subjects gave their informed and written consent to participate in the study protocol. All subjects were investigated both at...
sea level and after ascent by car and cable car to the Angelo Mosso Institute at Col d'Olen (2,950 m altitude), Gressoney La Trinité, Italy.

At sea level and at a moderately high altitude, BGM reliability at different blood glucose levels was tested, and blood glucose was assessed in fasting and resting conditions at 7:00 A.M.; at 10:00 A.M., before an in-field exercise test; and immediately, 5 min, and 15 min after the exercise stopped. Capillary glucose was simultaneously assessed with the OT and the GE. Both of these BGMs measure capillary blood glucose through the glucose oxidase-peroxidase reaction. BGMs were calibrated at the beginning of each test session. A venous blood sample was simultaneously drawn from the contralateral antecubital vein in a sodium fluoride tube, centrifuged, and stored at −20°C. Plasma glucose was assayed with the glucose oxidase method (GO) within 3 days. This last assessment was taken as a reference method. Statistical analysis compared BGM capillary glucose values and GO plasma glucose values for each blood collection time. Measurement linearity was tested with Pearson's correlation coefficient. The mean of the differences between the BGM and GO results represents the mean bias between the methods with accuracy expressed as percent error (PE):

$$PE(\%) = \frac{BGM - GO}{GO} \times 100\%$$

The level of statistical significance was considered to be $P < 0.05$.

The GE and OT measurements had a good correlation with plasma glucose both at moderately high altitude and at sea level. Pearson's correlation coefficients were 0.960 and 0.981 for the GE and 0.946 and 0.985 for the OT at sea level and at moderately high altitude, respectively. Biases between plasma glucose and BGM measurements were as follows: for the GE, $-5.9 \pm 27.3$ at sea level and $4.9 \pm 21.6$ at moderately high altitude; for the OT, $4.5 \pm 32.3$ at sea level and $-13.3 \pm 22.7$ at moderately high altitude. At sea level, both the GE and the OT tended to underestimate glucose values (NS); at moderately high altitude, the GE tended to underestimate glucose values (NS); and the OT tended to underestimate glucose values (NS). Mean PEs between plasma glucose and BGM measurements were 0.5 (OT) and 4.1 (GE) at sea level and 6.4 (OT) and 8.1 (GE) at moderately high altitude. PE tended to be higher for both BGMs at moderately high altitude (NS). Figures 1 and 2 show the bias between the single measurements with both devices at sea level and at moderately high altitude, respectively. At moderately high altitude (Fig. 2), the tendency of the GE to overestimate was more evident for low (≤100 mg/dl) and intermediate (100–200 mg/dl) blood glucose values, whereas the OT tended to underestimate mainly high blood glucose values (NS).

In our study, BGM performance was similar and good at sea level. At a moderately high altitude, a tendency to overestimate blood glucose for the GE and to underestimate for the OT was observed. The overestimation for the GE involved mainly low (≤100 mg/dl) and intermediate (100–200 mg/dl) blood glucose values. This could present a problem in the presence of symptoms suggesting hypo-
glycemia and normal blood glucose values. The OT tended to underestimate mainly high blood glucose values, although its performance with low to intermediate values was good. The present study assessed the accuracy of two BGMs at a moderately high altitude in which changes in temperature, humidity, and PO$_2$ can result in errors in blood glucose determination (2). Our results are consistent with previous studies (2,3). The decrease in PO$_2$ could alter the second phase of the chromogen reaction and underestimate blood glucose values (4); on the other hand, an increase in atmospheric pressure could overestimate blood glucose values (5). In our study, minimal overestimation by the GE at low intermediate blood glucose values at moderately high altitude cannot be explained by the altered PO$_2$. An increase in hematocrit, which is known to alter blood glucose measurements with BGMs (6), may also occur after prolonged exposure to high altitude or as a consequence of dehydration. Although our study did not determine hematocrit, the exercise test was short, and the patients were instructed to drink according to their thirst during the 3,000-m exposure; therefore, dehydration was not likely to have occurred. In conclusion, BGM performance is similar and good at sea level. At a moderately high altitude similar to that experienced during winter skiing or summer hiking, a tendency to overestimate low to normal blood glucose values for the GE and to underestimate high blood glucose values for the OT was observed. The bias is not clinically meaningful for either BGM, both of which can be safely used by diabetic patients during exposure to moderately high altitudes. Some care in the evaluation of low and intermediate blood glucose values measured with the GE is nevertheless recommended.

**REFERENCES**


**COMMENTS AND RESPONSES**

**Deterioration of Glycemic Control After Long-Term Treatment With Troglitazone in Nonobese Type 2 Diabetic Patients**

Troglitazone is an oral antidiabetic drug used to treat type 2 diabetic patients with insulin resistance. Troglitazone improves overall insulin sensitivity in the liver and skeletal muscles, which are the largest consumers and metabolizers of glucose in the body (1-3). Recent reports showed that troglitazone is also effective in nonobese type 2 diabetic patients whose hyperglycemia could not be controlled with sulfonylurea therapy (4,5). However, we have been aware that in some patients in whom adequate glycemic control is obtained during the first several months of troglitazone treatment, their glycemic control deteriorates several months later. We assume that two distinct groups of type 2 diabetic patients exist who respond differently to long-term administration of troglitazone, one group that maintains a steady response and another group that has a decreasing response after several periods. In this study, we retrospectively examined 20 patients with type 2 diabetes who were treated with troglitazone for ≥12 months and whose HbA$_1c$ levels had improved by ≥1% with troglitazone by month 6.

In 8 of the 20 patients (40%), HbA$_1c$ levels increased by ≥0.5% after 6-9 months despite continuous troglitazone treatment (group P). In contrast, the rest of the patients experienced steady glycemic control with <0.5% of HbA$_1c$ fluctuation (group G). During the first 5 months, HbA$_1c$ levels decreased from means ± SEM 9.1 ± 0.9 to 7.3 ± 0.8% in group P and from 8.5 ± 1.2 to 6.8 ± 0.8% in group G, respectively. No significant differences were evident between the two groups regarding the decrease in HbA$_1c$ during the first 5 months (Fig. 1). From month 6 onward, HbA$_1c$ levels in group P climbed gradually by 0.2% a month up to the baseline level at month 12, but HbA$_1c$ levels were stable in group G throughout the treatment period. A significant difference in HbA$_1c$ levels was evident during months 6-12 (P < 0.05).

Among clinical characteristics, group P had a significantly lower BMI (22.5 ± 3.2 vs. 26.7 ± 4.1 kg/m$^2$, P < 0.05) and significantly lower fasting insulin levels (4.9 ± 3.6 9 vs. 9.3 ± 4.9 μU/mL, P < 0.05). Of the 10 patients with a BMI of <25 kg/m$^2$ (80%), 8 exhibited deterioration of glycemic control.

In this study, we report a group of patients who showed a renewed decline in glycemic control after long-term treatment with troglitazone. These results seemed to suggest a secondary failure of troglitazone. Our study demonstrates that this drug is indeed useful for a long-standing obese insulin-resistant diabetes but not for a nonobese type 2 diabetes.

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References


Figure 1—Change from baseline in HbA1c. Values are means ± SEM.