Fibroblast Growth Factor 21 Levels in Young Healthy Females Display Day and Night Variations and Are Increased in Response to Short-Term Energy Deprivation Through a Leptin-Independent Pathway

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\textbf{OBJECTIVE}—Fibroblast growth factor (FGF)-21 is an endocrine factor with potent metabolic effects. Its day–night patterns of secretion and/or its physiological response to energy deprivation and relationship to free fatty acids (FFA) and/or leptin remain to be fully elucidated. We aim to elucidate day–night pattern of FGF-21 levels and its relationship to FFA, to assess whether energy deprivation alters its circulating patterns, and to examine whether leptin may mediate these changes.

\textbf{RESEARCH DESIGN AND METHODS}—Six healthy lean females were studied for 72 h in a cross-over interventional study under three different conditions: on isocaloric diet and in a fasting state with administration of either placebo or metreleptin in physiological replacement doses. Blood samples were obtained hourly from 8:00 A.M. on day 4 until 8:00 A.M. on day 5.

\textbf{RESULTS}—FGF-21 exhibited day–night variation pattern during the isocaloric fed state. Fasting significantly increased FGF-21 levels ($P < 0.01$) via a leptin-independent pathway. Day–night variation pattern in the fed state was lost on fasting. Leptin replacement in the hypoleptinemic state restored approximate entropy of FGF-21 time series but did not alter circulating levels. FGF-21 levels were closely cross-correlated with FFA levels in all three states.

\textbf{CONCLUSIONS}—A day–night variation in the levels of FGF-21 exists in young lean females in the fed state. Energy deprivation increases FGF-21 levels via a leptin-independent pathway. The interaction between FGF-21 and starvation induced lipolysis as indicated by its close cross-correlations with FFA in both fed state and energy deprivation needs to be studied further.

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Received 14 March 2012 and accepted 4 September 2012.  
DOI: 10.2337/dc12-0497. Clinical trial reg. no. NCT00140231, clinicaltrials.gov  
This article contains Supplementary Data online at http://care.diabetesjournals.org/lookup/suppl/doi:10.2337/dc12-0497/-/DC1.  
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Leptin is an adipokine with a pivotal role in signaling energy availability in the energy-deficient state (15). Energy-deficient states, leading to hypoleptinemia, induce several neuroendocrine adaptations facilitating the mobilization of alternative energy sources via processes such as lipolysis (15,16). FGF-21 levels have been previously shown to be elevated in subjects with anorexia nervosa, a condition characterized by hypoleptinemia (17). Moreover, circulating levels of FGF-21 have been shown to be strongly related to leptin levels in both anorectic and normal weight women (18). We have previously shown that decreasing leptin levels mediate some of these neuroendocrine adaptations to starvation in mice (19) and humans (15,20,21). However, it is unknown if an interaction exists between leptin and FGF-21.

In this study, we aimed to examine the relationship between FFA and FGF-21, and to elucidate the biological rhythm of FGF-21 in both energy-replete and energy-deficient states. We also aimed to determine whether an interaction exists between FGF-21 and leptin in view of previous studies highlighting a relationship between FGF-21 and energy homeostasis. These studies present a comprehensive examination of the biological characteristics of FGF-21 and clarify the relationship between lipolysis and FGF-21 in humans, thereby shedding light on the role of FGF-21 in energy homeostasis and diabetes in humans, and thus paving the way on how future clinical studies of FGF-21 can be interpreted.

**RESEARCH DESIGN AND METHODS**—Six young, healthy, and lean women (age, 22.8 ± 3.4 years; BMI, 21.7 ± 2.2 kg/m²) who were eumenorrheic were enrolled in a clinical research center–based, randomized, cross-over interventional study involving three separate 5-day-long inpatient admissions (22). Six subjects with a cross-over design, enabling paired comparisons, would provide 80% power to detect a difference of 1.4 SD between different conditions at the conventional α=0.05 level. In the first admission, the subjects were studied in the isocaloric fed state, whereas in the following two admissions the subjects were studied in the prolonged fasting state for 72 h and were randomized to receive either placebo or metreleptin at replacement doses. A cross-over to the opposite arm took place in the later admission so that all six subjects received both placebo and metreleptin. The subjects were admitted on day 1 at 9:00 P.M. the night before the commencement of the study on day 2. The study concluded after the 3-day intervention and ended on day 5, when they were discharged after the last blood-taking at 8:00 A.M. the study admissions were separated by at least 8 weeks to enable adequate washout and recovery of metabolic status. All subjects had a regular menstrual cycle and were not using any medications, including oral contraceptive pills. Study visits were standardized to occur between the days 6 and 11 of their menstrual cycles.

In the fed-state study, subjects were given a standardized isocaloric diet with breakfast at 8:00 A.M., lunch at 1:00 P.M., dinner at 6:00 P.M., and a snack at 10:00 P.M. daily. Caloric intake was distributed with 20% of calories from breakfast, 35% from lunch, 35% from dinner, and 10% from the evening snack.

In the prolonged fasting studies, subjects received only caffeine-free and calorie-free liquids for 3 days, which included NaCl (500 mg), KCl (40 mEq), and a standard multivitamin daily. Starting at 8:00 A.M. on day 1 of the fasting/leptin admission, metreleptin was administered as a subcutaneous injection every 6 h for 3 days, at a dose of 0.08 mg/kg per day on day 1 and 0.2 mg/kg per day on days 2 and 3, on the basis of previous pharmacokinetic studies (23–25). During the fasting/placebo admission, a buffer solution of similar volume was administered subcutaneously every 6 h, similar to the leptin arm.

All physical activities, light–dark intervals, and blood sampling schedule were standardized for all three studies. On day 3, blood was drawn through an indwelling intravenous catheter every 15 min from 8:00 A.M. on day 3 until 8:00 A.M. on day 4, and then was pooled every hour to meet the assays’ volume requirements.

The study protocols were approved by the Institutional Review Board of Beth Israel Deaconess Medical Center, and written informed consent was obtained from all the subjects. Clinical-quality metreleptin (formally known as r-metHuLeptin) was supplied by Amgen Inc. (Thousand Oaks, CA) and administered under an investigational new drug application submitted to the Food and Drug Administration.

**Assays**

Serum FGF-21 was measured by ELISA (R&D Systems, Minneapolis, MN), with a sensitivity of 4.67 pg/mL, intra-assay coefficient of variation of 2.9–3.9%, and interassay coefficient of variation of 5.2–10.9%, in accordance with the manufacturer’s instructions. All serum samples were stored at −80°C until analysis and were analyzed in duplicate. Leptin and insulin levels were measured as previously reported (22). FFA levels were measured by an enzymatic colorimetric assay (Wako Diagnostic USA). Glucose was measured by an automated analyzer (Hitachi cobas c311; Roche Diagnostics).

**Statistical analysis**

Statistical analysis was performed using Stata version 12 (Stata Corp, College Station, TX). Descriptive statistics are presented as means ± SD. Normality of the variables was evaluated using the Shapiro-Wilk test. Variables that were not normally distributed were normalized using the appropriate, for each time, transformation. Analysis for the existence of any potential day–night variation pattern in FGF-21 and FFA levels and cross-correlation analysis between FGF-21 and FFA circulating levels were performed, at the level of each individual, using the COSINE and CORRELATION algorithms of the Pulse XP software accordingly (UVA Pulse Analysis Software, Charlottesville, VA). In addition, we performed trigonometric ordinal least-squares (OLS) regression analysis on the data from all subjects, estimating the parameters of FGF-21 oscillations and the adjusted coefficient of determination (R²). Comparisons between mean FGF-21 and FFA levels across the three states were performed with hierarchical mixed-effects linear models. The 24-h trajectories of the analytes were expressed as a linear function of time at the level 1 of the model, and “state” was introduced as a level 2 predictor using dummy encoding. The optimal model was selected based on Akaike information criterion (AIC) and Bayesian information criterion (BIC). Normality and homoscedasticity of residuals and random effects were verified through frequency histograms and box plots. Area under the curve (AUC) of FGF-21 was estimated using the trapezoid method and correlations of the change of FGF-21 AUC and the change at the levels of different hormones were analyzed using simple linear regression. Comparisons between baseline levels and AUC across the three groups were performed with repeated-measures ANOVA because of the cross-over design of the study. Significant comparisons were further analyzed with post hoc paired comparisons using the least significant difference correction for multiple comparisons.
The adjusted coefficient of determination \( (R^2) \) that reflects the percent of the variability that is explained by an underlying day–night pattern is very high \( (R^2 = 48.28\% ; \ P < 0.001) \). This enables us to report with great confidence that there is clinically important day–night variability. Clinically important day–night secretion patterns traditionally have been associated with adjusted coefficients of determination \( >15\% \). As an example, studies of cortisol, a hormone with well-established circadian patterns of secretion, have revealed an \( R^2 \) of \( >30\% \) in similar models with the same sample size (27).

**Energy deprivation state with 72-h fast and placebo replacement**

Serum leptin levels decreased to <20% of baseline in response to energy deprivation (14.66 to 2.78 ng/mL; \( P < 0.001 \)). Normalized AUC of FGF-21 levels were significantly higher during energy deprivation compared with the fed state (\( P = 0.0066 \)) (Fig. 2A). Linear regression analysis revealed no association between FGF-21 levels at 8:00 a.m. and FGF-21 AUC in the fasting state (\( \beta = 0.56 ; \ P = 0.25 \)). OLS regression analysis revealed absence of any clinically significant day–night variation pattern of circulating FGF-21 levels during the energy deprivation state. Similar to FGF-21 levels, circulating FFA levels do not exhibit any clinically significant day–night variation pattern while in the fasted state (adjusted coefficient of determination 9.31%).

**Energy deprivation state with 72-h fast and lepton replacement**

Leptin replacement restored leptin levels to normal physiological range as previously reported (22), but had no effect on circulating FGF-21 levels. FGF-21 AUC during leptin replacement remained significantly higher compared with the isocaloric fed state (\( P = 0.0079 \)) and was not statistically different from the AUC in the fasting state (\( P = 0.9165 \)) (Fig. 2A). Similar to the fasting/placebo arm, OLS regression analysis revealed absence of any clinically significant day–night variation pattern of circulating FGF-21 and FFA levels.

**ApEn**

During fasting, the ApEn of the FGF-21 time series decreased from 0.731 ± 0.096 to 0.588 ± 0.114 [Tukey honestly significant difference (HSD) corrected for multiple comparisons, \( P < 0.05 \)], whereas leptin replacement therapy restored ApEn back to 0.712 ± 0.102 (Tukey HSD corrected for multiple comparisons, \( P < 0.05 \)) (Fig. 2B).

**Relationship between changes in FGF-21 AUC with other hormones/substrates during fasting**

Other hormones and substrates of interest, including insulin, FFA, and glucose, were analyzed to elucidate possible association with the increase of FGF-21 during energy deprivation. There were no correlations between changes in insulin (\( P = 0.539 \)), glucose (\( P = 0.12 \)), and the increase in FGF-21 levels after fasting. However, FGF-21 AUC increased with a quadratic relationship to the increase in FFA AUC after fasting (standardized regression coefficients: linear \( \beta = -2.31 \); quadratic \( \beta = 3.22 \); adjusted \( R^2 = 99.79\%; \ P < 0.001 \)) (Fig. 2D); individuals who exhibited the largest increase in the FFA levels from the fed to the fasting state also exhibited the largest increase in the FGF-21 levels.

**Relationship of FGF-21 with FFA**

FGF-21 levels were cross-correlated with FFA levels both in the fed and the fasting states (Fig. 3). Cross-correlation analysis demonstrated that the 24-h circulating pattern of FGF-21 was closely associated to the FFA 24-h pattern in both fasting and fed states. While the subjects were in the isocaloric fed state, four out of six subjects exhibited significant positive cross-correlation ranging from 0.35 to 0.81 between FGF-21 and FFA levels at 2- to 6-h lag, demonstrating that high levels of FFA were followed by high levels of FGF-21 with a 2- to 6-h lag (Fig. 3). Similarly, during the fasted state, in four out of six subjects FGF-21 and FFA levels were positively cross-correlated (0.25–0.52) at 5- to 8-h lag, demonstrating that high levels of FFA were followed by high levels of FGF-21 with a 5- to 8-h lag. At the high level of FGF-21, FFA was noted to momentarily decrease before recovering to high levels subsequently in both fed and fasting states. Regarding the subjects who did not exhibit significant cross-correlation, there were two subjects in the fed and two subjects in the fasting condition. One of these subjects was the same in the fed and fasting conditions. The other one subject who did not exhibit cross-correlation was different between the fed and fasting states.

**CONCLUSIONS**—We report herein, using a controlled interventional study design, that FGF-21 levels display a
Fasting-induced FGF-21 increase observed here is in agreement with data from Galman et al. (9) reporting no change in FGF-21 levels after a 2-day fast or feeding of a ketogenic diet. Similarly, Christodoulides et al. (13) reported no significant variation in FGF-21 levels during 48-h fasting followed by a 24-h refeeding. Yu et al. (12) reported a significant increase in FGF-21 levels during overnight fasting, although the increase diminished at the end of a 24-h fast. If the postulation that the day–night variation of FGF-21 demonstrated herein is related to overnight energy deprivation-induced lipolysis, then we hypothesize that FGF-21 levels should be increased in association with adequate fasting-induced lipolysis. This study clearly demonstrates an increase in FGF-21 in response to 72 h of fasting, i.e., a period clearly longer than that in previous studies. Besides the increase in its levels with fasting, the day–night variation pattern of FGF-21 shown in the fed state was abolished. Similar responses in both the placebo and leptin arms in this study reinforce the reproducibility of the FGF-21 increase in response to fasting.

The response of FGF-21 levels to fasting remains unclear to date. Galman et al. (9) reported no change in FGF-21 levels after a 2-day fast or feeding of a ketogenic diet. Similarly, Christodoulides et al. (13) reported no significant variation in FGF-21 levels during 48-h fasting followed by a 24-h refeeding. Yu et al. (12) reported a significant increase in FGF-21 levels during overnight fasting, although the increase diminished at the end of a 24-h fast. If the postulation that the day–night variation of FGF-21 demonstrated herein is related to overnight energy deprivation-induced lipolysis, then we hypothesize that FGF-21 levels should be increased in association with adequate fasting-induced lipolysis. This study clearly demonstrates an increase in FGF-21 in response to 72 h of fasting, i.e., a period clearly longer than that in previous studies. Besides the increase in its levels with fasting, the day–night variation pattern of FGF-21 shown in the fed state was abolished. 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reported by Galman et al. (9) that demonstrated that prolonged fasting of 7 days with significant ketosis increased FGF-21 levels. The discrepancy in relation to other studies could be secondary to the shorter duration of fasting (11,13), different assays used (10–12,13), different genders studied (13), and design of these studies. In particular, we found a poor correlation between a single early morning determination of FGF-21 and the total FGF-21 levels over the course of a 24-h period as determined by the AUC. Therefore, studies utilizing a single determination of FGF-21 level at a specific time point may not represent FGF-21 physiology fully (10,12,13). It is also possible that a sex dimorphism might exist in FGF-21 physiology, considering that a sex difference exists in lipids metabolism (30). These questions remain to be fully addressed by future studies in both genders in the fed and fasting states.

The increase of FGF-21 in response to fasting indicates a possible role of this molecule in mediating some of the metabolic adaptations in energy deprivation. Studies in rodents have demonstrated an increase in FGF-21 levels in fasting induced directly by peroxisome proliferator-activated receptor-α in liver (31,32). Peroxisome proliferator-activated receptor-α regulates the utilization of fat as an energy source during starvation and is the molecular target for drugs used to treat dyslipidemia. These studies in rodents identify hepatic FGF-21 as a regulator of lipid homeostasis and highlight a physiological role in adaptation to a low-energy state for this hepatic hormone. We herein demonstrate a positive relationship between the increase in FFA in response to fasting and the increase in FGF-21 levels, with subjects having higher FFA levels also having a greater increase in FGF-21 levels (Fig. 2C). Moreover, FFA levels were observed to precede the peaking of FGF-21 by a few hours, suggesting that the increase in FFA might have a role in inducing the peaking of FGF-21. This is supported by previous observations in which the physiologically elevated FFAs induced by lipid–heparin infusion were reported to increase circulating FGF-21 levels in humans (33). Interestingly, apart from the observed increase in FGF-21 levels that followed after the peaks of FFA, FFA levels were noted to decrease momentarily after the peak of FGF-21 levels in this study, suggesting a possible feedback interaction between FFA and FGF-21. This observation is consistent with recent studies demonstrating that FGF-21 may play a role in inhibiting hormone-stimulated lipolysis in human and murine adipocytes (34,35). Although causality remains to be proven by future interventional studies, the present findings highlight the possibility that FGF-21...
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Figure 3—A: Mean FGF-21 and FFA for all six subjects in the fed state. Solid lines represent FGF-21 levels and interrupted lines represent FFA levels (n = 6). B: Mean FGF-21 and FFA for all six subjects in the fasting with placebo state. Solid lines represent FGF-21 levels and interrupted lines represent FFA levels (n = 6). C: Mean FGF-21 and FFA for all six subjects in the fasting with leptin replacement state. Solid lines represent FGF-21 levels and interrupted lines represent FFA levels (n = 6). (A high-quality color representation of this figure is available in the online issue.)
by demonstrating that FGF-21 displays day–night variation pattern in the fed state and is increased in energy deprivation via a leptin-independent mechanism. We also show for the first time that leptin replacement restored the approximate entropy of FGF-21 time series, but not the energy deprivation-induced changes of FGF-21 levels. Finally, we demonstrate that the day–night variation and the increase of FGF-21 production in response to fasting are closely related to FFA levels. The knowledge of these biological characteristics of FGF-21 is critical for future clinical studies to plan the timing and the situations in which samples are collected to evaluate FGF-21 levels across different individuals or groups with comparable results. Furthermore, given the significant role of lipolysis in insulin resistance and diabetes, the relationship between FGF-21 and lipolysis elucidated in this study paves the way on how future studies on FGF-21 and diabetes can be interpreted. Further studies are necessary to replicate these data in men, given the gender dimorphism in lipids metabolism (30), and to delineate the precise interactions between FGF-21 and all other hormones and substrates involved in energy homeostasis to clarify the metabolic role and clinical applications of FGF-21 in humans.

Acknowledgments—This study was supported by the National Institute of Diabetes and Digestive and Kidney Diseases grants 58785, 79929, and 81913. The project described also was supported by Award Number 1I01CX000422-01A1 from the Clinical Science Research and Development Service of the VA Office of Research and Development. Amylin Pharmaceuticals, Inc. supplied metetlpep in for this study but had no role in the study design, conduct of the study, collection, management, analysis, and interpretation of the data, or the preparation, review, or approval of the manuscript. Funding was also received from the National Institutes of Health National Center for Research Resources grant M01-RR-01032 (Harvard Clinical and Translational Science Center). The content is solely the responsibility of the authors and does not necessarily represent the official views of the National Center for Research Resources or the National Institutes of Health.

No potential conflicts of interest relevant to this article were reported.

J.-P.F. wrote the manuscript, researched data, and performed laboratory work. K.N.A. wrote the manuscript and analyzed data. J.P.C., J.P., and H.-S.M. performed laboratory work. C.S.M. is the principal investigator and reviewed and edited the manuscript. C.S.M. is the guarantor of this work and, as such, had full access to all of the data in the study and takes responsibility for the integrity of the data and the accuracy of the data analysis.

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