QUANTITATIVE ESTIMATION OF INSULIN SENSITIVITY IN TYPE 1 DIABETIC SUBJECTS WEARING A SENSOR AUGMENTED INSULIN PUMP

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ABSTRACT

Objective
The goal was to develop a new index of insulin sensitivity in patients with type 1 diabetes estimated from continuous glucose monitoring (CGM) and subcutaneous insulin delivery data under carefully controlled conditions.

Research Design & Methods
The data base consists of 12 subjects with type 1 diabetes, studied during breakfast, lunch and dinner, in a clinical research unit, wearing both subcutaneous insulin pump and CGM. Frequent blood samples were drawn for measurements of plasma glucose and insulin concentrations in order to estimate insulin sensitivity with the oral minimal model (\( S_{I}^{MM} \)). The new index of insulin sensitivity (\( S_{I}^{SP} \)) was calculated with a simple algebraic formula, for each meal, using only CGM and insulin pump data and compared with \( S_{I}^{MM} \).

Results
\( S_{I}^{SP} \) was well correlated with \( S_{I}^{MM} \) (\( r = 0.825, p<10^{-8} \)) and diurnal pattern was also similar to \( S_{I}^{MM} \).

Conclusion
A novel method for estimating insulin sensitivity in subjects with type 1 diabetes on sensor augmented insulin pump therapy has been presented. This new index correlates well with the reference oral minimal model estimate of insulin sensitivity. The knowledge of patient-specific insulin sensitivity and its diurnal variation can help in optimizing insulin therapy in type 1 diabetes and could also inform the next generation closed-loop control systems.
The standard therapy for type 1 diabetes consists of exogenous insulin administration, either by multiple daily injections or with continuous subcutaneous insulin infusion (CSII) through the insulin pump, adjusted according to self-monitored blood glucose (SMBG) levels 3-4 times a day. However, in the last 10-15 years, new possibilities in diabetes therapy have emerged thanks to continuous glucose monitoring (CGM) and CSII, which substitute SMBG and multiple daily injection therapy respectively. Minimally-invasive CGM devices can measure, in real-time, interstitial glucose concentrations in continuous time for up to several days. CSII uses the subcutaneous route and administer insulin with a basal/bolus strategy, i.e. continuously deliver insulin over 24 hours and inject boluses.

In order to determine the appropriate meal insulin bolus, it would be important to know the subject-specific insulin sensitivity, i.e. the ability of insulin to stimulate glucose utilization and inhibit glucose production. Indices of insulin sensitivity include those based on hyperinsulinemic euglycemic clamp [1], IVGTT [2] and more recently, meal (MTT) and oral glucose tolerance test (OGTT). These indices include the oral glucose minimal model [3][4][5], a minimal model based integral formula [5] and surrogate measures [6][7][8]. However, all the above require the measurement of both plasma glucose and insulin concentrations, and therefore cannot be used in everyday life. In the outpatient setting, the only usable approach is the risk-method proposed by Breton and Kovatchev [9] which uses SMBG data collected over a period of 2-6 weeks and some patient parameters. This index measures the average insulin sensitivity in the preceding 2 weeks, and thus cannot be used to assess the intraday variability of insulin sensitivity.

Here we propose a method to estimate insulin sensitivity from CGM sensor and insulin pump data and validate it against the oral minimal model, which uses plasma glucose and insulin concentrations. The method is usable in everyday life in patients wearing the two devices.

RESEARCH DESIGN & METHODS
Data Base and Protocol

Twelve type 1 diabetic subjects (5 females, age 39.5±14.2 years, BMI 25.7±3.8 kg/m², HbA₁C ≤ 8.5 % or 69 mmol/mol) were studied for three days in the Clinical Research Unit of the Mayo CTSA as part of a separate study to determine diurnal patterns of insulin sensitivity [10]. Briefly, once a day, a triple-tracer mixed meal study protocol was performed during breakfast, lunch or dinner in latin square design. Blood samples were collected at -180, -30, 0, 5, 10, 20, 30, 60, 90, 120, 150, 180, 240, 300, 360, with t = 0 corresponding to meal time, for measurement of plasma glucose and insulin concentrations in order to estimate $S_I$ with the oral minimal model [3], here considered as reference. More details can be found in [10]. Subjects also wore both subcutaneous insulin pump (Medtronic or Insulet OmniPod®) and continuous glucose monitoring (Dexcom Seven® Plus). Within subject mean absolute relative difference (MARD) between CGM readings and reference was equal to 12.1±3.3% and no systematically variation over time was found (mean standard deviation within subjects was 5.3±3.7%). This is perfectly in line with published reports [11]. Figure 1 shows CGM and insulin infusion rate in a representative subject. The above 12 subjects have been chosen among the 19 reported in [10] since they had the complete CGM and CSII data required for the calculation described below. Of note one CGM trace was missing in one subject during one meal.

The Basis

The starting point is the derivation of insulin sensitivity by integrating the oral minimal model equations [3], as described in Caumo et al. [5]. Then, the calculation is adapted to allow the use of CGM and CSII data, instead of plasma glucose and insulin concentrations. The oral minimal model [3] is:
\[
\begin{align*}
\dot{G}(t) &= -[p_1 + X(t)] \cdot G(t) + p_1 \cdot G_b + \frac{Ra_G(t)}{V_G} \\
\dot{X}(t) &= -p_2 \cdot X(t) + p_3 \cdot [(t) - l_b] \\
G(0) &= G_b \\
X(0) &= 0
\end{align*}
\]

(1)

where \( G \) is plasma glucose concentration [mg/dL], with \( G_b \) denoting its basal value, \( X \) insulin action [min\(^{-1}\)], \( I \) plasma insulin concentration [\( \mu \)U/mL], with \( I_b \) denoting its basal value, \( Ra_G \) post-hepatic appearance of meal glucose [mg/kg/min], \( V_G \) glucose distribution volume [dL/kg], \( p_2 \) [min\(^{-1}\)] speed of rise and decay of insulin action and \( p_3 \) [min\(^{-2}\) per \( \mu \)U/mL] its size. Insulin sensitivity is defined as:

\[
S_i = \frac{p_3}{p_2} \cdot V_G
\]

(2)

By substituting \( X(t) = \frac{p_3}{p_2} \cdot X'(t) \) and \( p_1 = \frac{GEZI}{V_G} + \frac{S_i}{V_G} \cdot I_b \) into Eq. 1 we obtain:

\[
\begin{align*}
\dot{G}(t) &= -\left[ \frac{GEZI}{V_G} + \frac{S_i}{V_G} \cdot I_b + \frac{S_i}{V_G} \cdot X'(t) \right] \cdot G(t) + \left( \frac{GEZI}{V_G} + \frac{S_i}{V_G} \cdot I_b \right) \cdot G_b + \frac{Ra_G(t)}{V_G} \\
\dot{X}(t) &= -p_2 \cdot X'(t) + p_3 \cdot [(t) - l_b] \\
G(0) &= G_b \\
X(0) &= 0
\end{align*}
\]

(3)

By integrating the differential equation (Eq. 3) from the time of the meal ingestion (\( t_{meal} \)) to the end of the experiment (\( t_{end} \)) and rearranging, one has:

\[
S_i = \frac{\int_{t_{meal}}^{t_{end}} Ra_G(t) dt - GEZI \int_{t_{meal}}^{t_{end}} \Delta G(t) dt - V_G \int_{t_{meal}}^{t_{end}} G(t) dt}{\int_{t_{meal}}^{t_{end}} X'(t) \cdot G(t) dt + I_b \int_{t_{meal}}^{t_{end}} \Delta G(t) dt}
\]

(4)
The first integral in the numerator can be rewritten as

$$\int_{t_{meal}}^{t_{end}} Ra_g(t) dt = \frac{D \cdot f(t_{end})}{BW} \quad (5)$$

where $D$ [mg] is the amount of glucose ingested during the meal and $f(t_{end})$ the fraction of the ingested dose which at $t=t_{end}$ has reached plasma. $X^*$ in Eq. 4 is usually not available because model parameters are unknown, thus the denominator of Eq. 4 is substituted with the average of glucose excursion times the overall insulin stimulus:

$$\frac{1}{(t_{end} - t_{meal})} \int_{t_{meal}}^{t_{end}} f(t) dt \cdot \int_{t_{meal}}^{t_{end}} |G(t)| dt \quad (6)$$

where $|\Delta G|$ is above basal plasma glucose concentration.

Thus $S_I$ is given by:

$$S_I = \frac{D \cdot f(t_{end}) - GEZI \cdot AUC(\Delta G) - V_G \cdot [G(t_{end}) - G(t_{meal})]}{AUC(\Delta G) \cdot \frac{AUC(\Delta G)}{t_{end} - t_{meal}}} \quad (7)$$

where AUC is the area under the curve calculated from the start of the meal ($t_{meal}$) to the end of the experiment ($t_{end}$). Subject’s specific parameters used by the formula are the body weight (BW) [kg], height [m] and age [years]. BW is explicitly used in Eq. 7 and, together with height and age, employed for the calculation of plasma insulin clearance (CL), as discussed below. Parameter fixed to population values are glucose effectiveness at zero insulin GEZI [dL/kg/min] (fixed to 0.01
dL/kg/min for diabetic subjects [10][18]) and volume of glucose distribution $V_G$ [dL/kg] (fixed to 1.45 dL/kg, according to [3]).

$G$, $\Delta G$ and $I$ are not directly available but can be derived from CGM and subcutaneous insulin delivery as detailed below.

**Glucose signal from CGM**

CGM measures the interstitial glucose concentration (IG) which is related to plasma glucose by a first order differential equation, i.e. IG is a delayed version of plasma glucose:

$$
\begin{aligned}
\begin{cases}
\dot{I}_G(t) &= -k \cdot I_G(t) + k \cdot G(t) \\
CI_{GM}(t) &= I_G(t)
\end{cases}
\end{aligned}
$$

(8)

Thus, with a well-calibrated device, assuming $I_G(t_{end})=I_G(t_{meal})$, one has

$$
\int_{t_{meal}}^{t_{end}} \dot{I}_G(t) dt = 0 = -k \int_{t_{meal}}^{t_{end}} I_G(t) dt + k \int_{t_{meal}}^{t_{end}} G(t) dt
$$

(9)

i.e.

$$
AUC(G) = AUC(I_G) = AUC(CGM)
$$

(10)

Similarly, for the above basal glucose signal one has:

$$
AUC(\Delta G) = AUC(\Delta CGM)
$$

(11)
Thus one can safely assume that, in presence of well-calibrated device, AUC(ΔCGM) and AUC(|ΔCGM|) are good approximations of AUC(ΔG) and AUC(|ΔG|), respectively.

In presence of a non-calibrated device, if at least two SMBG samples are available, it is possible to off-line recalibrate the CGM profile [12][13].

**Insulin signal from CSII**

For the calculation of AUC(I), we assume that insulin is not degraded locally in the site of infusion. In this case, the integral of plasma insulin can be obtained from subcutaneous insulin infusion divided by the plasma insulin clearance CL. In fact, plasma insulin kinetics can be described with a single compartment model [14]:

\[
I(t) = -n \cdot I(t) + \frac{R_a(t)}{V_i}
\]

where \(I(t)\) is the plasma insulin concentration, \(R_a(t)\) the insulin rate of appearance in plasma, \(V_i\) the insulin volume of distribution and \(n\) the fractional insulin clearance rate \((n = CL/V_i)\).

Then, integrating Eq. 12 from the time of the pre-meal bolus \(t_{meal}\) to the end of the observation period \(t_{end}\) and assuming that insulin is back to its initial value at the end of the experiment, one has:

\[
0 = -n \int_{t_{meal}}^{t_{end}} I(t) dt + \int_{t_{meal}}^{t_{end}} \frac{R_a(t)}{V_i} dt
\]

Finally, under the assumption that all infused insulin eventually reaches the circulation, one has:
Thus we can compute $AUC(I)$ from the amount of insulin infused subcutaneously and $CL$:

$$AUC(I) = \frac{1}{CL} \int_{t_{meal}}^{t_{end}} \text{basal}(t)dt + \sum_{t_k=t_{meal}}^{t_{end}} \frac{\text{bolus}(t_k)}{CL}$$

(15)

where $\text{basal}(t)$ is the basal insulin infusion rate during the integration period, $\text{bolus}(t_k)$ the pre-meal or correction bolus administered at $t=t_k$, $CL$ [L/min] plasma insulin clearance, which can be calculated from subject’s height, BW and age, by using the population model proposed in [17].

In addition, if correction boluses are administered before the start of the meal, one has to consider that part of that injected insulin could be still active. The residual active insulin can be determined by adopting the same algorithm presented in [15] which uses the Insulin on Board (IOB) curves adapted from [16]. This quantity ($\text{IOB}(t_{meal})$) must be added to the previously estimated $AUC(I)$. Moreover, if correction boluses are administered before the end of the considered interval, IOB is used to evaluate the active insulin at the end of the study ($\text{IOB}(t_{end})$), which is subtracted from the previously estimated $AUC(I)$. In summary:

$$AUC(I) = \frac{1}{CL} \int_{t_{meal}}^{t_{end}} \text{basal}(t)dt + \sum_{t_k=t_{meal}}^{t_{end}} \frac{\text{bolus}(t_k)}{CL} + \text{IOB}(t_{meal}) - \text{IOB}(t_{end})$$

(16)

**Accounting for carbohydrates on board (COB)**

As evident from Eq. 7, an accurate calculation of insulin sensitivity requires the knowledge of the amount of carbohydrates (CHO) entering the circulation in the integration interval ($D\cdot f(t_{end})$). In this study, meals were provided at 7 am, 1 pm and 7 pm in each day, thus time interval between the
meals was of at least 6 hours. However, this may not always be the case. Furthermore, not all the
CHO ingested with a meal may be fully absorbed before the ingestion of a second meal. Thus, to
account for unabsorbed CHO, e.g. frequent meals close to each other, the concept of carbohydrates
on board (COB) is introduced. Similarly to IOB, COB is a function which, at each time t, quantifies
the fraction of the ingested CHO that has not yet appeared in the circulation. COB is based on the
model of gastrointestinal tract [19]: given the amount of carbohydrates ingested at time \( t_m \), COB provides, at each time \( t > t_m \), the percentage of carbohydrates not yet absorbed, while for \( t > t_m > 360 \)
min, it is assumed that the carbohydrates absorption is almost completed. The fraction of the
ingested dose which has reached plasma at time t, \( f(t) \), can be calculated as the ratio between the
AUC of the meal rate of appearance and the ingested dose, \( D \) (Figure 2, left panel), assuming that,
at the end of the meal, the fraction of the meal appearing in plasma is equal to \( f_\infty = 0.9 \) (Figure 2,
right panel). The time course of \( f \) is shown in Figure 2, right panel. COB is then calculated as
\[
COB(t) = f_\infty - f(t).
\]
To better grasp the use of \( f(t) \) and COB(t) for the calculation of insulin sensitivity, let us consider
the following example: suppose that a subject eats 50 g of CHO at \( t=0 \) min and another 40 g at
\( t=180 \) min, then fasts for more than 360 min. One can calculate two values of \( S_I \), one for each meal
ingestion (\( S_I^{\text{meal1}} \) and \( S_I^{\text{meal2}} \)). For the calculation of \( S_I^{\text{meal1}} \) the amount of ingested glucose to be
considered is \( 50 \cdot f(180) \) g of CHO, while for the calculation of the second \( S_I^{\text{meal2}} \), one should use
\( 50 \cdot \text{COB}(180) + 40 \cdot f_\infty \).
Generalizing, the amount of CHO (AoC) to use in the formula for the i-th meal is:
\[
AoC^{\text{meal i}} = D(t_{\text{meal i}}) \cdot f(t_{\text{end i}}) + \text{COB}(t_{\text{end i-1}}) \cdot D(t_{\text{meal i-1}})
\]  
(17)
where \( D(t_{\text{meal i}}) \) is the amount of CHO ingested at the time of the i-th meal \( (t_{\text{meal i}}) \), \( t_{\text{end i}} \) the time at
which the i-th meal ends (corresponding to the time of ingestion of the \((i + 1)-th\) meal), \( \text{COB}(t_{\text{end i-1}}) \)
the carbohydrates on board at the end of the \((i-1)\)-th meal which contained \(D(t_{meal}^{i-1})\) amount of carbohydrates.

**Insulin Sensitivity from Sensor and Pump Data \((S_{I}^{SP})\)**

Incorporating the above derivations into Eq. 7 one can estimate insulin sensitivity from sensor and pump data \((S_{I}^{SP})\) for the \(i\)-th meal [20]:

\[
S_{I}^{SP}(meal^{i}) = \frac{AOC(meal^{i})}{BW} - GEZI \cdot AUC(\Delta CGM) - V_{G} \cdot [CGM(t_{end}^{i}) - CGM(t_{meal}^{i})] - \left[ \int_{t_{meal}^{i}}^{t_{end}^{i}} \frac{basal(t)}{CL} \, dt + \sum_{k=1}^{t_{end}^{i}} \frac{bolus(t_{k})}{CL} + IOB(t_{meal}^{i}) - IOB(t_{end}^{i}) \right] \cdot \frac{AUC(\Delta CGM)}{(t_{end}^{i} - t_{meal}^{i})}
\]

\(18\)

It is important to define the domain of validity of Eq.18. When CGM is still high six hours after meal ingestion \(S_{I}^{SP}\) can become negative. This brings the method working outside its domain of validity. We thus recommend to use the formula only if the recalibrated \(\Delta CGM\) is lower than 150 mg/dL 6 hours after meal ingestion. As a matter of fact, one of our subjects (#4) during lunch has recalibrated \(\Delta CGM\) greater than 150 mg/dL 6 hour after meal ingestion, and \(S_{I}^{SP}\) was negative. This subject was thus excluded in the following analysis.

**Minimal Model Insulin Sensitivity \((S_{I}^{MM})\) and Validation of \(S_{I}^{SP}\)**

The oral glucose minimal model [3][4] was used to estimate insulin sensitivity, from plasma glucose and insulin concentrations, in twelve subjects studied three times [10]. Here we consider these measures as reference values \((S_{I}^{MM})\) to which \(S_{I}^{SP}\) are compared for validation.

**Assessment in Case of Non-Completely Absorbed Meals**
In our experimental protocol meals were well spaced (at least 6 h) and thus completely absorbed before the next meal. However, in daily life this may not happen consistently. Thus, it is important to assess the performance of the method in case of non-completely absorbed meals. We did so by comparing estimates of $S_I^{SP}$ obtained with a 360 min interval to those obtained from shorter intervals (up to 180 min), both with and without employing the COB function.

**Reproducibility**

When proposing a new metric like $S_I^{SP}$, it is important to assess its reproducibility, i.e. if it provides similar values when repeatedly applied to the same subject under the same experimental conditions. However, due to the large intra-subject variability of insulin sensitivity [10], $S_I^{SP}$ will likely change if the same meal is administered in two occasions to a subject. Thus, to address reproducibility one can resort to simulation. We evaluated $S_I^{SP}$ reproducibility in simulation using our FDA accepted T1D simulator [21]. We simulated a 7-days scenario for 100 in silico subjects with 3 meals per day (breakfast: from 6AM to 8AM, lunch: from 11.30AM to 1.30PM, dinner: from 6PM to 8.30PM) with different amounts (breakfast: 0.7÷0.9 g/kg, lunch: 0.8÷1.0 g/kg, dinner: 0.8÷1.3 g/kg), while subject-specific insulin sensitivity was maintained constant for the whole simulation. A total of 21 $S_I^{SP}$ were thus calculated for each subject (3 values per 7 days). To assess the repeatability of the index, we calculated the average (mean), the standard deviation (SD) and the coefficient of variation (CV=SD/mean) of the 21 estimates of $S_I^{SP}$, in each subject.

**Statistical Analysis**

Data are presented as mean ± SD. Two sample comparisons were done by Paired Sample t-test. Pearson’s correlation was used to evaluate univariate correlation.
RESULTS

The correlation between the two indices was very good ($r = 0.825$, $p<10^{-8}$; Figure 3, right panel) and diurnal pattern was similar, indicating that, apart from a scale factor, $S_I^{SP}$ closely mirrors $S_I^{MM}$. $S_I^{MM}$ and $S_I^{SP}$ have been estimated in the twelve subjects at breakfast, lunch and dinner. $S_I^{SP}$ was significantly higher than $S_I^{MM}$ (13.86±14.56 vs. 6.67±5.63 dL/kg/min per µU/mL, $p<10^{-3}$; Figure 3, left panel).

When $S_I^{SP}$ was calculated for non-completely absorbed meals, i.e. relying on reduced integration intervals, mean values of $S_I^{SP}$ were virtually the same (Figure 4, top left) if one employs the COB function. Conversely, $S_I^{SP}$ increased systematically if COB is not used (Figure 4, top right). Of note, the correlation between $S_I^{SP}$ calculated at the end of the experiment and that obtained from reduced integration intervals decreases only slightly (Figure 4, middle panels) and that the absolute relative error increases (Figure 4, bottom panels) both with and without employing COB. Finally, in silico reproducibility of $S_I^{SP}$ was 23±6%.

DISCUSSION

Insulin sensitivity is a key parameter of the metabolic status of an individual which could be beneficial also for optimizing insulin therapy in type 1 diabetes. In fact, the knowledge of patient specific $S_I$ and its daily variation can truly help in determining the optimal insulin bolus to be administered to cover the ingested carbohydrates. However, all methods available for the estimation of insulin sensitivity rely on plasma glucose and insulin measurements and thus cannot be used in everyday life of a patient with type 1 diabetes.

In this paper we have proposed an index of insulin sensitivity, $S_I^{SP}$, which can be estimated in patients with type 1 diabetes wearing a CGM sensor and an insulin pump. We have demonstrated that it is similar to the one obtained with the oral minimal model, which requires plasma glucose
and insulin data. The method uses retrospective subcutaneous sensor and insulin delivery data with some anthropometric parameters for each subject, and provides, for each meal, patient’s insulin sensitivity by an integral formula.

$S_{SP}$ measures how subcutaneously infused insulin affects the CGM profile. Thus, $S_{SP}$ is not exactly the same index derived with the minimal model ($S_{MM}$), which represents the ability of insulin to suppress endogenous glucose production and stimulate glucose uptake. In other words, $S_{SP}$ is a new metric of insulin sensitivity with its own range. In fact, mean values of $S_{SP}$ were almost twice $S_{MM}$. However, the correlation between the two indices was excellent ($r = 0.825, p<10^{-6}$).

A robust estimate of insulin sensitivity can be obtained for each meal, whenever meals are well spaced (5-6 h). However, we also tested the method in case of non-completely absorbed meals. To deal with this situation, we introduced the concept of the Carbohydrates on Board (COB). Similarly to Insulin on Board [16], COB represents, at each time $t$, the amount of ingested glucose which has not yet been absorbed. Thus, one can define different COB curves for different types of meal, i.e. fast or slow carbohydrates. We used COB in the $S_{SP}$ calculation to evaluate the correct amount of carbohydrates in relation with the observed CGM profiles in a given time interval. We demonstrated the need of employing COB: in case of a short time interval between consecutive meals, a robust estimation of $S_{SP}$ can only be obtained if carbohydrates absorption is taken into account (Figure 4, top left vs. top right panel).

Possible applications of the new index include its use for assessing intra- and inter-day variability (e.g. existence of diurnal patterns) of insulin sensitivity in large cohorts of subjects with type 1 diabetes, in normal life conditions. For instance, the method is currently being applied to the STAR 3 data of type 1 diabetic subjects wearing a sensor augmented insulin pump [22]. This will provide, in each subject, the $S_{SP}$ time course during several months and will allow testing for the existence of subject-specific $S_{SP}$ daily patterns and correlation of such variation with lifestyle and other factors. Following validation using STAR 3 data, a clinical application would be the use of $S_{SP}$ to calculate the optimal insulin to carbohydrate ratio (CR) [23][24]. Thus, optimizing CR based on
recent CGM and CSII data collected 1-2 weeks prior to a clinical visit may be useful for physicians to improve patient-specific meal bolus insulin therapy and thus a major component of glucose control. In addition, the knowledge of patient CR daily pattern may help in the design of optimal closed-loop control algorithms relying on patient-specific open-loop insulin therapy at the time of transition from open to closed loop therapy. A natural progression of this work would be the development of real time adaptation of open or closed loop therapy based on the presence of well-developed metrics such as the one developed here. Further development of diabetes technology hardware and algorithms should make this a reality over the next several years.

The method relies on data provided by CGM device which can occasionally suffer from inaccuracies. For instance, the assumption that AUC(CGM) is a good approximation of AUC(G) becomes critical if the device is not well calibrated. If this occurs S$_{1}^{SP}$ will also reflect sensor inaccuracy. However, to improve the quality of CGM measurements, some algorithms can be used to recalibrate CGM traces [12][13]. Another possible limitation is the need to fix some parameters (GEZI and V$_G$) to population values [10][18][3] and others calculated from population models (CL) using anthropometric data [17]. In order to test the effect of fixing these parameters, we also calculated S$_{1}^{SP}$ using individualized GEZI, estimated with the oral glucose minimal model [3] and CL, directly estimated from the data in each patient: we obtained values very similar to those obtained with fixed parameters, and a slightly higher correlation with S$_{1}^{MM}$.

**CONCLUSIONS**

Insulin sensitivity is an important element in the daily life of patients with type 1 diabetes and could be useful to optimize insulin therapy. However, methods to estimate this index by emerging technologies, such as subcutaneous CGM sensor and insulin pump, has never been proposed. We have presented a method which estimates insulin sensitivity from CGM and insulin pump data.
Future studies involving a larger data bases that include larger cohorts of subjects studied for a longer time, are needed to better define the applicability in free living conditions.

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Michele Schiavon, Chiara Dalla Man and Claudio Cobelli developed the method, analyzed the results and drafted the manuscript. Dr. Ananda Basu and Dr. Yogish Kudva commented and edited the manuscript. The algorithm proposed in this paper is part of a patent request deposited by the University of Padova (No. 13/661,755). Claudio Cobelli is the guarantor of this work and, as such, had full access to all the data in the study and takes responsibility for the integrity of the data and the accuracy of the data analysis.

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AUTHOR DISCLOSURE STATEMENT

There are no conflicts of interest to declare for any of the authors.
FIGURE LEGEND

Figure 1: Data needed for the estimation of insulin sensitivity. Glucose data are measured by CGM (left panel, black line), with some SMBG data available for sensor recalibration (left panel, gray dots). Insulin data are the basal infusion and boluses administered by insulin pump (right panel, black line and gray triangles respectively).

Figure 2: Time course of meal rate of appearance, $Ra_G$ (left), and fraction of meal which appears in plasma, $f$ (right panel).

Figure 3: Mean values (left panel) and correlation plot (right panel) between $S_{IMM}$ and $S_{ISP}$ insulin sensitivity indices for breakfast, lunch and dinner (white, gray and black bars). * indicates $p<10^{-3}$ with paired sample $t$-test.

Figure 4: Sensitivity analysis of $S_{ISP}$ with (left panel) and without (right panel) accounting for carbohydrates on board for different time integration intervals: mean values of $S_{ISP}$ (top), correlation indices (middle), and absolute relative error (bottom), calculated with respect to $S_{ISP}$, i.e. estimated at $t = 360$ min.
REFERENCES


Figure 1

71x28mm (600 x 600 DPI)
Figure 2

74x31mm (600 x 600 DPI)
Figure 3

68x26mm (600 x 600 DPI)
Figure 4