

Oxidative Stress and Insulin Resistance

The Coronary Artery Risk Development in Young Adults study

KYONG PARK, PHD¹
MYRON GROSS, PHD²
DUK-HEE LEE, MD³
PAUL HOLVOET, PHD⁴

JOHN H. HIMES, PHD¹
JAMES M. SHIKANY, DRPH⁵
DAVID R. JACOBS, JR., PHD^{1,6}

OBJECTIVE — Although cumulative evidence suggests that increased oxidative stress may lead to insulin resistance in vivo or in vitro, community-based studies are scarce. This study examined the longitudinal relationships of oxidative stress biomarkers with the development of insulin resistance and whether these relationships were independent of obesity in nondiabetic young adults.

RESEARCH DESIGN AND METHODS — Biomarkers of oxidative stress (F₂-isoprostanes [F₂Isop] and oxidized LDL [oxLDL]), insulin resistance (the homeostasis model assessment of insulin resistance [HOMA-IR]), and various fatness measures (BMI, waist circumference, and estimated percent fat) were obtained in a population-based observational study (Coronary Artery Risk Development in Young Adults) and its ancillary study (Young Adult Longitudinal Trends in Antioxidants) during 2000–2006.

RESULTS — There were substantial increases in estimated mean HOMA-IR over time. OxLDL and F₂Isop showed little association with each other. Mean evolving HOMA-IR increased with increasing levels of oxidative stress markers ($P < 0.001$ for oxLDL and $P = 0.06$ for F₂Isop), measured in 2000–2001. After additional adjustment for adiposity, a positive association between oxLDL and HOMA-IR was strongly evident, whereas the association between F₂Isop and HOMA-IR was not.

CONCLUSIONS — We observed positive associations between each of two oxidative stress markers and insulin resistance. The association with oxidized LDL was independent of obesity, but that with F₂Isop was not.

Diabetes Care 32:1302–1307, 2009

Clinical type 2 diabetes is considered to be preceded by a long period of insulin resistance, during which blood glucose is maintained at near-normal levels by compensatory hyperinsulinemia (1). Convincing evidence has established that the level of insulin resistance is a pre-diabetic state that can predict incident type 2 diabetes relatively far into the future (2).

Increased oxidative stress appears to be a deleterious factor leading to insulin

resistance, β -cell dysfunction, impaired glucose tolerance, and, ultimately, type 2 diabetes (3,4). Obesity may play a role in the relationship between systemic oxidative stress and these conditions (5). Chronic oxidative stress is particularly dangerous for β -cells because pancreatic islets are among those tissues that have the lowest levels of antioxidant enzyme expression, and β -cells have high oxidative energy requirements (4). In addition, there is considerable evidence that in-

creased free radicals impair glucose-stimulated insulin secretion, decrease the gene expression of key β -cell genes, and induce cell death (4,6). If β -cell functioning is impaired, it results in an underproduction of insulin, fasting hyperglycemia, and, eventually, the development of type 2 diabetes (7).

However, most previous studies investigating this association have been in vitro or in small in vivo studies (8,9), and data presenting an association between the degree of oxidative stress and the risk of developing insulin resistance among nondiabetic people in the community are scarce (9). We explored the associations between oxidative stress and insulin resistance to see whether elevated levels of oxidative stress markers increase the risk of future insulin resistance, whether different biomarkers of oxidative stress show consistent results, and whether these associations can be explained by obesity in a longitudinal design in a population-based cohort. We elected to study F₂-isoprostanes (F₂Isop) (10) and oxidized LDL (oxLDL) (11–13), which mark complementary areas of systemic oxidative stress. To strengthen our hypothesized direction from oxidative stress to insulin resistance, we tested the reciprocal relationship to see if elevated levels of HOMA-IR were associated with an increase in the level of one oxidative stress marker longitudinally.

RESEARCH DESIGN AND METHODS

Data from the Coronary Artery Risk Development in Young Adults (CARDIA) study and its ancillary study, Young Adult Longitudinal Trends in Antioxidants (YALTA), were used to examine the association between biomarkers of oxidative stress and insulin resistance. Briefly, CARDIA is a longitudinal study aiming to investigate lifestyle and other factors that influence the evolution of cardiovascular disease in young adults. This study began in 1985 with a cohort of 5,115 healthy black and white men and women, aged 18–30 years, who were free-living individuals residing in four U.S. cities (Birmingham, AL; Chicago, IL; Minneapolis, MN; and Oakland, CA). The YALTA ancillary study analyzes blood

From the ¹Division of Epidemiology and Community Health, School of Public Health, University of Minnesota, Minneapolis, Minnesota; the ²Department of Laboratory Medicine and Pathology, University of Minnesota, Minneapolis, Minnesota; the ³Department of Preventive Medicine, School of Medicine, College of Medicine, Kyungpook National University, Daegu, Korea; the ⁴Atherosclerosis and Metabolism Unit, Katholieke Universiteit Leuven, Leuven, Belgium; the ⁵Division of Preventive Medicine, University of Alabama at Birmingham, Birmingham, Alabama; and the ⁶Department of Nutrition, University of Oslo, Oslo, Norway.

Corresponding author: David R. Jacobs, jacobs@epi.umn.edu.

Received 10 February 2009 and accepted 10 April 2009.

Published ahead of print at <http://care.diabetesjournals.org> on 23 April 2009. DOI: 10.2337/dc09-0259.

© 2009 by the American Diabetes Association. Readers may use this article as long as the work is properly cited, the use is educational and not for profit, and the work is not altered. See <http://creativecommons.org/licenses/by-nc-nd/3.0/> for details.

The costs of publication of this article were defrayed in part by the payment of page charges. This article must therefore be hereby marked "advertisement" in accordance with 18 U.S.C. Section 1734 solely to indicate this fact.

and urine biomarkers of oxidative stress, endothelial dysfunction, and related concepts and interprets those data in conjunction with the other data that have been collected by CARDIA. Follow-up examinations were conducted during 1987–1988 (year 2), 1990–1991 (year 5), 1992–1993 (year 7), 1995–1996 (year 10), 2000–2001 (year 15), and 2005–2006 (year 20). The percentages of the surviving cohort who returned for these examinations were 90, 86, 81, 79, 74, and 72%, respectively. In this analysis, we included all participants with F_2 Isop or oxLDL measurements at year 15 and fasting insulin/fasting glucose measurements at year 20. Among them, we excluded those who had been diagnosed with diabetes, defined as a fasting glucose ≥ 126 mg/dl (7 mmol/l) or who were receiving antidiabetes medication between years 0 and 20 in our analysis.

Self-reported demographic (age, sex, race, and educational level) and behavioral (smoking, alcohol consumption, and physical activity) information was obtained across CARDIA examination visits. Educational status was quantified as the reported number of years of schooling completed. Self-reported alcohol beverages were quantified as average milliliters of alcohol consumed per day, and smoking status was classified as never, former, or current smoker. An interviewer-administered questionnaire queried participation spent in leisure, occupational, and household physical activities over the course of a year, weighting frequency and intensity in order to obtain a total activity score.

Anthropometry

The participants wore light clothes without shoes during anthropometry. Body weight was recorded to the nearest 0.2 pound. Body height was recorded to the nearest 0.5 cm, and BMI was calculated as weight in kilograms divided by the square of height in meters, using the average of the two measurements. Waist circumference (cm) was measured with each participant's weight distributed equally on both feet. The measurement was made midway between the iliac crest and the lowest lateral portion of the ribcage and anteriorly midway between the xyphoid process of the sternum and the umbilicus, keeping the tape (vinyl anthropometric centimeter tape) horizontal.

Since the adiposity measurements, total fat mass, and percent fat were obtained only from a subsample ($n = 1,072$)

of the study participants, race- and sex-specific predictive equations for percent body fatness (%FAT) were computed using the year 10 dual-energy X-ray absorptiometry (DXA) examination (14); these estimates enabled study of total body fatness of each participant. Year 10 DXA was obtained only from a subsample ($n = 1,072$), in Birmingham and Oakland, of the study participants using the QDR 2000 (Hologic, Bedford, MA) with software version 11.1. These race-/sex-specific predictive equations included age, BMI, waist circumference (referred to as WC), and squared terms for waist circumference and BMI (WC^2 and BMI^2): for black men: %FAT = $-55.5371 + 0.0578 \times \text{age} + 0.1585 \times \text{BMI} + 1.1741 \times \text{WC} - 0.00365 \times \text{WC}^2$; for black women: %FAT = $-52.0734 + 0.0901 \times \text{age} + 1.9772 \times \text{BMI} + 0.9664 \times \text{WC} - 0.015 \times \text{BMI}^2 - 0.0051 \times \text{WC}^2$; for white men: %FAT = $-82.6174 + 0.1015 \times \text{age} + 0.5448 \times \text{BMI} + 1.5883 \times \text{WC} - 0.0065 \times \text{WC}^2$; and for white women: %FAT = $-75.2741 + 0.1698 \times \text{age} + 1.2678 \times \text{BMI} + 1.7154 \times \text{WC} - 0.00972 \times \text{WC}^2$. These equations had higher correlation coefficients and smaller mean square errors and predictive errors compared with the widely used general predictive equations for body composition, such as the Durnin and Womersley equations (15). The correlations of observed year 10%FAT with predicted values were $R^2 = 0.66$ for black men, 0.76 for black women, 0.60 for white men, 0.74 for white women, and 0.82 for sex and race combined, compared with $R^2 = \sim 0.75$ for the Durnin and Womersley equations that relied solely on skinfolds. These formulae closely predicted DXA measured fat and lean mass at year 20.

Blood components

Before drawing blood, all of the participants were asked to fast for 12 h and to abstain from smoking for 2 h before their examinations. After venous blood samples were drawn, plasma was separated by centrifugation, transferred into airtight vials, stored at -70°C , and shipped on dry ice.

Fasting glucose and insulin were measured at years 0, 7, 10, 15, and 20. Glucose was measured in stored blood samples using the hexokinase ultraviolet method on a Cobas Mira Plus chemistry analyzer. The insulin measurements were performed by using a radioimmunoassay with an overnight, equilibrium-incubation

format. Based on reassays of glucose in December 2007 in ~ 200 samples stored since year 7 and 100 samples stored since year 15, glucose and insulin were recalibrated to harmonize them with the previous measurements. Recalibrated glucose values were $6.98 + 0.94 \times \text{year 7 glucose concentration}$, $7.15 + 0.96 \times \text{year 10 glucose concentration}$, $6.99 + 1.01 \times \text{year 15 glucose concentration}$, and $4.06 + 0.97 \times \text{year 20 glucose concentration}$. Recalibrated insulin was $-0.36 + 0.93 \times \text{year 20 insulin concentration}$.

The biomarkers of oxidative stress were obtained at year 15 and/or year 20. F_2 Isop was measured at years 15 and 20 with gas chromatography–mass spectrometry. OxLDL concentrations were measured by the Mercodia oxidized LDL ELISA (Mercodia, Uppsala, Sweden) in year 15 samples that had been stored at -70°C for several years. The antibody (mAb-4E6) is directed against a conformational epitope in the apolipoprotein B-100 moiety of LDL, as a consequence of substitution of ≥ 60 lysine residues of apolipoprotein B-100 with aldehydes, and these substituting aldehydes can be produced by peroxidation of lipids of LDL, leading to the generation of oxLDL. This method was compared with another assay, developed by Holvoet et al. (11), and showed the similar analytical performance and clinical applicability (11).

Plasma levels of ascorbic acid were measured using high-performance liquid chromatography (HPLC) at year 15. Levels of carotenoids (including lycopene, α -carotene, β -carotene, β -cryptoxanthin, and zeaxanthin plus lutein) and tocopherols (α -tocopherol and γ -tocopherol) were obtained by HPLC at year 15. A common protocol and quality-control procedures were used for all examinations.

Statistical analysis

BMI and waist circumference were set to missing at each examination at which a woman was pregnant. We investigated patterns of repeated measurements for individuals with large within-person SDs from year 0 to year 20 by visual inspection of the raw data to detect if there were any substantial departures (e.g., outliers) from patterns. Six outliers of waist circumference at year 15, one outlier of BMI at year 20, one outlier of F_2 Isop at year 15, and one outlier of HOMA-IR at year 7 were replaced with missing values. Insulin resistance was assessed by HOMA-IR, which was calculated as fasting plasma insulin (mU/l) \times fasting plasma glucose

(mmol/l)/22.5 (16). HOMA-IR, F₂Isop, oxLDL, and circulating antioxidant (ascorbic acid, lycopene, α -carotene, β -carotene, β -cryptoxanthin, zeaxanthin plus lutein, α -tocopherol, and γ -tocopherol) estimates were logarithmically transformed, since their distributions were skewed to the right. The subjects were classified into quartiles, based on their levels of oxidative stress markers. Partial correlation coefficients between body fat measurements/antioxidants and oxidative stress markers were calculated by sex, adjusting for age and race.

Multivariable generalized linear models were used to test associations of oxidative stress markers with evolving HOMA-IR, using as dependent variable HOMA-IR at year 20 and adjusting for demographic and lifestyle factors (e.g., age, sex, race, study center, smoking status, physical activity, alcohol consumption, and education) and the previous measure of HOMA-IR at year 15. In addition, the models were adjusted for adiposity measurements (e.g., BMI, waist circumference, and equation-driven estimates of percent fat) to examine if the association between oxidative stress and insulin resistance was independent of obesity. All analyses were conducted using SAS version 9.1.

RESULTS — The mean age of the participants during 2000–2001 (year 15) was 40 years, and 45% were men among 2,774 nondiabetic study participants. Participants who had higher levels of F₂Isop were more likely to be women ($P < 0.001$) (Table 1). In contrast, women had lower levels of oxLDL ($P < 0.001$). Education showed an inverse association with both markers ($P < 0.001$). An increased level of F₂Isop was positively associated with smoking ($P < 0.001$), whereas oxLDL was not associated with this factor. Physical activity was inversely associated with both markers, though the gradient was stronger over F₂Isop than over oxLDL.

Body fat measurements showed strong positive associations with both markers ($P < 0.01$), as shown in Table 1. F₂Isop showed much stronger correlations with adiposity markers in women than in men (Table 2). Correlations between F₂Isop and body fat measurements, including BMI, waist circumference, and estimated body percent fat, were 0.40–0.43 in women, whereas these correlations were only 0.15–0.16 in men, after adjustment for race and age. OxLDL did

not show a sex difference in the correlations with body fat measurements. F₂Isop was negatively correlated with sum of five serum carotenoids, ascorbic acid, and α -tocopherol but positively correlated with serum γ -tocopherol. In contrast, the correlations between oxLDL and antioxidants were minimal. Interestingly, the two oxidative stress markers, Isop and oxLDL, were minimally correlated in our sample ($r = 0.09$, $P = 0.0007$, in women and $r = -0.003$, $P = 0.9$, in men).

Table 3 includes adjusted estimates for year 20 HOMA-IR, according to the quartiles of year 15 F₂Isop and oxLDL. We observed a significant increase in the estimates of evolving HOMA-IR with increasing levels of oxLDL ($P < 0.001$) and a marginally significant association with F₂Isop ($P = 0.06$). After additional adjustment for the adiposity measurements, oxLDL was still significantly associated with year 20 HOMA-IR, whereas the association between F₂Isop and HOMA-IR became substantially attenuated. Further adjustments for HDL cholesterol, LDL cholesterol, and triglycerides did not affect the magnitude and significance of the association between oxLDL and HOMA-IR. Adiposity measurements remained significant, predicting HOMA-IR in the models adjusted for either marker of oxidative stress ($P < 0.0001$).

In our analysis, we did not adjust for medication use (e.g., lipid- or blood pressure-lowering medications) because such use could mediate or be directly correlated with mediators in the causal pathway for associations of oxidative stress on insulin resistance. Omission of those taking these medications did not materially change our study results (data not shown). There were no significant interactions between oxidative stress markers and other covariates, including sex, race, smoking, adiposity measurements, and physical activity. For example, although the association between F₂Isop and HOMA-IR was positive in whites and negative in blacks, race-specific coefficients were small and P values for interactions were not significant.

Increase in F₂Isop was not predicted by HOMA-IR in longitudinal analysis, controlling for body fat measurements (BMI, waist circumference, or estimated body percent fat), age, sex, race, study center, smoking status, physical activity, alcohol consumption, education, and F₂Isop, all of which were measured in the same year as the independent variable (BMI-adjusted regression coefficient =

-0.004 , $P = 0.9$; waist circumference-adjusted regression coefficient = -0.009 , $P = 0.7$; body fat percent-adjusted regression coefficient = -0.008 , $P = 0.7$).

CONCLUSIONS — We observed positive associations between oxidative stress markers and insulin resistance in a nondiabetic adult population. OxLDL was positively associated with insulin resistance, even after accounting for various adiposity measurements, a finding akin to a recent report (12) in these data that oxLDL predicted incident metabolic syndrome. Although the association between F₂Isop and insulin resistance was also positive, it was explained by additional adjustment for adiposity measurements. The levels of year 20 HOMA-IR predicted by each oxidative stress marker were not significantly different by sex. Recently, Meigs et al. (9) showed in a cross-sectional design that the association between HOMA-IR and oxidative stress, measured by urinary isoprostanes (8-epi-PGF 2 α /creatinine), was significant, independent of BMI, but association between insulin resistance prevalence and oxidative stress was attenuated by additional adjustment for BMI ($P = 0.06$) in individuals without type 2 diabetes in the Framingham Offspring Cohort Study (9). Our study substantially extends what is known about the impact of oxidative stress on insulin resistance in people without diabetes. In our longitudinal analysis, the results show that plasma F₂Isop was marginally related with elevated levels of HOMA-IR, but a substantial part of this association was explained by anthropometric measurements (BMI and waist circumference). Furthermore, we examined this association by using an alternate marker, oxLDL, that showed a stronger association with insulin resistance, independent of body fatness, compared with F₂Isop.

Oxidative stress results from an imbalanced condition in which the generation of free radicals is greater than the capacity of the antioxidant defense system to detoxify them (17). When oxidative stress is chronic, it is thought to result in damage to DNA, lipids, proteins, and other molecules, which may contribute to the development and progression of chronic disease, including cardiovascular disease and cancer (13,18). Recently, various indirect markers of oxidative stress have been used in epidemiological studies to measure oxidation damage due to fea-

Table 1—Means (or proportions) of demographic, lifestyle, and type 2 diabetes-related factors by quartile of oxidative stress markers*

2000–2001 variable (except where indicated)	Quartiles of year 15 F ₂ Isop				P _{trend}	Quartiles of year 15 oxLDL				P _{trend}
	1st quartile	2nd quartile	3rd quartile	4th quartile		1st quartile	2nd quartile	3rd quartile	4th quartile	
n	693	693	695	693		652	650	652	652	
Demographics										
Age (years)	40.3 ± 3.5	40.2 ± 3.5	40.2 ± 3.6	40.0 ± 3.7	0.2	40.1 ± 3.6	40.2 ± 3.6	40.2 ± 3.5	40.2 ± 3.7	0.9
White (%)	58.3	55.8	57.8	54.6	0.4	62.0	56.6	55.8	53.2	0.01
Men (%)	57.1	56.0	46.3	22.4	<0.0001	35.7	39.9	50.8	55.5	<0.0001
Education (grade of school completed)	15.3 ± 2.6	15.1 ± 2.5	15.0 ± 2.5	14.7 ± 2.4	<0.0001	15.4 ± 2.6	15.0 ± 2.5	15.0 ± 2.5	14.8 ± 2.4	<0.0001
Lifestyle										
Alcohol intake (g/day)	9.2 ± 24.8	10.3 ± 18.2	12.3 ± 24.0	13.1 ± 33.1	0.05	11.5 ± 21.5	9.9 ± 20.0	11.9 ± 31.3	12.0 ± 28.5	0.6
Physical activity (exercise units)	399 ± 295	394 ± 295	357 ± 286	286 ± 253	<0.0001	382 ± 289	354.4 ± 283	364 ± 292	339 ± 270	0.01
Current smoker (%)	14.5	19.5	21.9	25.4	<0.0001	19.2	20.4	19.2	22.0	0.5
Body fat components										
Waist circumference (cm)	85.4 ± 12.3	86.7 ± 11.8	87.7 ± 13.1	91.6 ± 14.3	<0.0001	81.9 ± 12.1	86.3 ± 13.1	89.4 ± 12.3	93.7 ± 12.0	<0.0001
BMI (kg/m ²)	26.2 ± 4.7	27.0 ± 4.7	27.9 ± 5.9	31.0 ± 7.2	<0.0001	25.9 ± 5.4	27.9 ± 6.4	28.5 ± 5.5	29.9 ± 5.8	<0.0001
Estimated body fat (%)	29.6 ± 8.6	30.7 ± 9.2	33.3 ± 10.1	40.3 ± 10.5	<0.0001	31.9 ± 9.9	33.9 ± 10.8	33.5 ± 10.6	34.5 ± 10.4	<0.0001
Oxidative stress markers										
ln (F ₂ Isop) (ng/l)	3.47 ± 0.2	3.81 ± 0.1	4.09 ± 0.1	4.58 ± 0.3	<0.0001	3.98 ± 0.5	4.00 ± 0.4	3.98 ± 0.5	4.0 ± 0.5	0.7
ln (oxLDL) (units/l)	4.32 ± 0.3	4.36 ± 0.3	4.32 ± 0.4	4.33 ± 0.3	0.7	3.89 ± 0.2	4.25 ± 0.1	4.47 ± 0.1	4.73 ± 0.1	<0.0001
ln (sum of five carotenoids) (μg/dl)†‡	4.74 ± 0.4	4.64 ± 0.4	4.55 ± 0.4	4.41 ± 0.4	<0.0001	4.63 ± 0.4	4.59 ± 0.4	4.58 ± 0.4	4.54 ± 0.4	0.0002
ln (ascorbic acid) (μg/dl)	2.29 ± 0.4	2.21 ± 0.4	2.15 ± 0.5	2.10 ± 0.5	<0.0001	2.22 ± 0.5	2.22 ± 0.4	2.16 ± 0.5	2.16 ± 0.4	0.02
ln (α-tocopherol) (μg/dl)‡	0.84 ± 0.1	0.80 ± 0.1	0.76 ± 0.1	0.73 ± 0.1	<0.0001	0.79 ± 0.2	0.78 ± 0.2	0.77 ± 0.2	0.78 ± 0.2	0.08
ln (γ-tocopherol) (μg/dl)‡	0.19 ± 0.1	0.21 ± 0.1	0.22 ± 0.1	0.24 ± 0.1	<0.0001	0.21 ± 0.1	0.21 ± 0.1	0.21 ± 0.1	0.22 ± 0.1	0.008
HOMA-IR (μU · l ⁻¹ · mmol ⁻¹ · l ⁻¹)										
ln (HOMA-IR) at year 0	1.09 ± 0.3	1.11 ± 0.3	1.11 ± 0.3	1.14 ± 0.3	0.0008	1.06 ± 0.3	1.12 ± 0.3	1.12 ± 0.3	1.15 ± 0.3	<0.0001
ln (HOMA-IR) at year 7	1.26 ± 0.4	1.26 ± 0.3	1.27 ± 0.3	1.33 ± 0.4	<0.0001	1.17 ± 0.3	1.28 ± 0.4	1.28 ± 0.3	1.38 ± 0.4	<0.0001
ln (HOMA-IR) at year 10	1.23 ± 0.3	1.26 ± 0.3	1.27 ± 0.4	1.33 ± 0.4	<0.0001	1.17 ± 0.3	1.26 ± 0.3	1.29 ± 0.4	1.36 ± 0.4	<0.0001
ln (HOMA-IR) at year 15	1.24 ± 0.4	1.27 ± 0.4	1.31 ± 0.4	1.40 ± 0.4	<0.0001	1.14 ± 0.3	1.28 ± 0.4	1.34 ± 0.4	1.46 ± 0.4	<0.0001
ln (HOMA-IR) at year 20	1.31 ± 0.4	1.35 ± 0.4	1.38 ± 0.4	1.43 ± 0.4	<0.0001	1.22 ± 0.4	1.34 ± 0.4	1.37 ± 0.4	1.52 ± 0.4	<0.0001

Data are means ± SD unless otherwise indicated. *All variables were measured at year 15, except as indicated. †Sum of five carotenoids: α-carotene, β-carotene, β-cryptoxanthin, lutein/zeaxanthin, and lycopene. ‡Adjusted for year 15 HDL cholesterol, LDL cholesterol, and triglycerides (25).

Table 2—Sex-specific partial correlation coefficients of observed body fat components, estimated percent body fat, and antioxidant markers at year 15 with F₂Isop and oxLDL

Adiposity and oxidative stress markers	F ₂ Isop				oxLDL			
	n	Women	n	Men	n	Women	n	Men
BMI (kg/m ²)*	1,509	0.43	1,260	0.15	1,418	0.25	1,184	0.23
Waist circumference (cm)*	1,506	0.40	1,259	0.16	1,416	0.31	1,183	0.26
Estimated body fat (%)‡	1,506	0.41	1,258	0.16	1,416	0.28	1,182	0.26
Sum of five carotenoids (μg/dl)*§	1,438	-0.40	1,196	-0.27	1,382	-0.02†	1,139	-0.06†
Ascorbic acids (μg/dl)*	1,255	-0.22	1,067	-0.24	1,206	-0.05†	1,017	0.004†
α-Tocopherol (μg/dl)*§	1,438	-0.29	1,196	-0.30	1,382	0.02†	1,139	-0.009†
γ-Tocopherol (μg/dl)*§	1,438	0.27	1,196	0.23	1,382	0.03†	1,139	-0.03†
F ₂ Isop (μg/dl)*	1,513	1	1,261	1	1,419	0.09	1,185	-0.003†

*All models were adjusted for age and race. †P > 0.05. ‡Based on race-/sex-specific prediction from BMI, waist circumference, and age; see RESEARCH DESIGN AND METHODS. §Additional adjustment for year 15 HDL cholesterol, LDL cholesterol, and triglycerides (25). ||Sum of five carotenoids: α-carotene, β-carotene, β-cryptoxanthin, lutein/zeaxanthin, and lycopene.

sibility and cost issues, instead of direct measures of free radicals and reactive oxygen species, which include electron resonance or spin trapping, capturing free radical reactions in real time (19). Among them, oxLDL and F₂Isop have been known as robust reflectors of oxidative stress in humans and have been widely used in biology, medicine, and epidemiological research (20,21). Each marker may capture different stages of the oxidative process or reflect different pathophysiological pathways (22). The apparent compartmentalization of oxidative stress is illustrated in the differing associations of F₂Isop, oxLDL, and different carotenoids with each other and with HOMA-IR. There was little correlation between the two measures of oxidative stress, though F₂Isop was well correlated with the sum of serum carotenoids and other serum antioxidant vitamins (e.g., ascorbic acid and α-tocopherol), lower values of which in part indicate reduced carotenoid intake and in part oxidative stress. In contrast, correlations between

oxLDL and these antioxidant nutrients were substantially weaker. In longitudinal analysis using CARDIA/YALTA, Hozawa et al. (23) reported that serum carotenoid concentrations were inversely associated with incident diabetes and HOMA-IR in nonsmokers, concluding that oxidative stress may be involved in the development of type 2 diabetes in nonsmokers. It has been reported that serum carotenoids were inversely associated with F₂Isop in the same data (24).

Our study shows that a substantial portion of the association between F₂Isop and HOMA-IR was explained by body fat measurements, and F₂Isop was strongly correlated with adiposity markers, especially in women. In a previous report using CARDIA/YALTA, Gross et al. (10) reported that the level of plasma F₂Isop was higher in women, especially for those with a BMI >25 kg/m², than men. Since F₂Isop is a product of lipid peroxidation, it is closely related to adipose tissue mass, especially for women. In contrast, correlations between oxLDL and body fat mea-

surements did not differ by sex. Furthermore, oxLDL was a strong precursor of the risk of insulin resistance, independent of obesity. Therefore, we suggest that each oxidative stress marker may capture a different oxidative stress pathway and that their interaction with adiposity differs between men and women. Nevertheless, in our study the estimated levels of evolving HOMA-IR with increasing levels of the oxidative stress markers did not differ significantly by sex.

Our study has several strengths. Studies examining the longitudinal associations between oxidative stress and insulin resistance are rare, especially with plasma oxidative stress markers in healthy young people in large, community-based studies, while most previous epidemiological studies have used urinary markers. Also, CARDIA has maintained a high participation rate and high-quality data collection through rigorous quality-control procedures throughout the study. Furthermore, we could test prediction in the reverse direction and added to the specificity of our finding by showing that HOMA-IR did not predict future oxidative stress, measured by F₂Isop, although we only measured oxLDL once and so were not able to measure the reverse direction with it.

Among potential limitations, first, oxidative stress is a highly complex phenomenon and our measures may not be sufficiently detailed to fully understand the role of oxidative stress and adipose tissue in development of insulin resistance. Second, the study was conducted only among black and white individuals, with no representation of Hispanic, Asian, or other individuals. As a result, the conclusions from this study may not be applicable to all populations. Finally,

Table 3—Relationship between year 15 oxidative stress markers and year 20 HOMA-IR

	F ₂ Isop		oxLDL	
	β-Coefficient ± SE	P	β-Coefficient ± SE	P
Model 1*	0.0244 ± 0.0158	0.1	0.0727 ± 0.0208	0.0005
Model 2†	0.0294 ± 0.0159	0.06	0.0753 ± 0.0208	0.0003
Model 3				
BMI (kg/m ²)‡	-0.0192 ± 0.0165	0.2	0.0598 ± 0.0206	0.004
Waist circumference (cm)‡	-0.0237 ± 0.0161	0.1	0.0432 ± 0.0206	0.04
Estimated body fat (%)‡	-0.0217 ± 0.0163	0.2	0.0501 ± 0.0206	0.02

β-Coefficient is a change in HOMA-IR [ln(mU · l⁻¹ · mmol⁻¹ · l⁻¹)] per one-unit increment of independent variables [ln(F₂Isop) (ng/l) or ln(oxLDL) (units/l)]. *Model 1: adjusting for age, sex, race, study center, and year 15 HOMA-IR. †Model 2: model 1 plus year 15 smoking status, physical activity, alcohol consumption, and education. ‡Model 3: model 2 plus year 15 body fat measurements (BMI, waist circumference, or estimated percent body fat; adding each variable separately).

there could be residual confounding due to this study's observational nature.

In summary, our results show that increased oxidative stress is associated with evolution of insulin resistance. The independent association of oxLDL with insulin resistance supports the hypothesis that oxidative stress leads to insulin resistance, independent of obesity. F₂Isop is closely correlated with adiposity, especially for women, and it is possible that F₂Isop is intermediate in a pathway that links insulin resistance with total body fat. Further research is needed to elucidate the underlying etiological relationships between adiposity and oxidative stress in the genesis and progression of insulin resistance by various biomarkers of oxidative stress and body fat measurements in diverse populations.

Acknowledgments— This research was supported by contracts N01-HC-95095, N01-HC-48047, N01-HC-48048, N01-HC-48049, N01-HC-48050, and grant R01-HL-53560—all from the National Heart, Lung, and Blood Institute, National Institutes of Health.

No potential conflicts of interest relevant to this article were reported.

References

1. Kahn SE. The relative contributions of insulin resistance and beta-cell dysfunction to the pathophysiology of type 2 diabetes. *Diabetologia* 2003;46:3–19
2. Martin BC, Warram JH, Krolewski AS, Bergman RN, Soeldner JS, Kahn CR. Role of glucose and insulin resistance in development of type 2 diabetes mellitus: results of a 25-year follow-up study. *Lancet* 1992;340:925–929
3. Evans JL, Goldfine ID, Maddux BA, Grodsky GM. Are oxidative stress-activated signaling pathways mediators of insulin resistance and β -cell dysfunction? *Diabetes* 2003;52:1–8
4. Shah S, Iqbal M, Karam J, Salifu M, McFarlane SI. Oxidative stress, glucose metabolism, and the prevention of type 2 diabetes: pathophysiological insights. *Antioxid Redox Signal* 2007;9:911–929
5. Davi G, Guagnano MT, Ciabattini G, Basili S, Falco A, Marinopicolini M, Nutini M, Sensi S, Patrono C. Platelet activation in obese women: role of inflammation and oxidant stress. *JAMA* 2002;288:2008–2014
6. Simmons RA. Developmental origins of diabetes: the role of oxidative stress. *Free Radic Biol Med* 2006;40:917–922
7. Kahn SE. The importance of the beta-cell in the pathogenesis of type 2 diabetes mellitus. *Am J Med* 2000;108:2S–8S
8. Ilyasova D, Morrow JD, Wagenknecht LE. Urinary F₂-isoprostanes are not associated with increased risk of type 2 diabetes. *Obes Res* 2005;13:1638–1644
9. Meigs JB, Larson MG, Fox CS, Keaney JF Jr, Vasan RS, Benjamin EJ. Association of oxidative stress, insulin resistance, and diabetes risk phenotypes: the Framingham Offspring Study. *Diabetes Care* 2007;30:2529–2535
10. Gross M, Steffes M, Jacobs DR Jr, Yu X, Lewis L, Lewis CE, Loria CM. Plasma F₂-isoprostanes and coronary artery calcification: the CARDIA Study. *Clin Chem* 2005;51:125–131
11. Holvoet P, Macy E, Landeloos M, Jones D, Jenny NS, Van de Werf F, Tracy RP. Analytical performance and diagnostic accuracy of immunometric assays for the measurement of circulating oxidized LDL. *Clin Chem* 2006;52:760–764
12. Holvoet P, Lee DH, Steffes M, Gross M, Jacobs DR Jr. Association between circulating oxidized low-density lipoprotein and incidence of the metabolic syndrome. *JAMA* 2008;299:2287–2293
13. Holvoet P, Jenny NS, Schreiner PJ, Tracy RP, Jacobs DR. The relationship between oxidized LDL and other cardiovascular risk factors and subclinical CVD in different ethnic groups: the Multi-Ethnic Study of Atherosclerosis (MESA). *Atherosclerosis* 2007;194:245–252
14. Yu X. *Predictive Equations of Body Fatness Based on DEXA Measurements*. Minneapolis, MN, University of Minnesota, 2003
15. Durnin JV, Womersley J. Body fat assessed from total body density and its estimation from skinfold thickness: measurements on 481 men and women aged from 16 to 72 years. *Br J Nutr* 1974;32:77–97
16. Matthews DR, Hosker JP, Rudenski AS, Naylor BA, Treacher DF, Turner RC. Homeostasis model assessment: insulin resistance and beta-cell function from fasting plasma glucose and insulin concentrations in man. *Diabetologia* 1985;28:412–419
17. Grune T. Oxidants and antioxidative defense. *Hum Exp Toxicol* 2002;21:61–62
18. Rossner P Jr, Gammon MD, Terry MB, Agrawal M, Zhang FF, Teitelbaum SL, Eng SM, Gaudet MM, Neugut AI, Santella RM. Relationship between urinary 15-F_{2t}-isoprostane and 8-oxodeoxyguanosine levels and breast cancer risk. *Cancer Epidemiol Biomarkers Prev* 2006;15:639–644
19. Vincent HK, Taylor AG. Biomarkers and potential mechanisms of obesity-induced oxidant stress in humans. *Int J Obes* 2006;30:400–418
20. Cracowski JL, Durand T, Bessard G. Isoprostanes as a biomarker of lipid peroxidation in humans: physiology, pharmacology and clinical implications. *Trends Pharmacol Sci* 2002;23:360–366
21. Tsimikas S. Measures of oxidative stress. *Clin Lab Med* 2006;26:571–590
22. Halliwell B, Whiteman M. Measuring reactive species and oxidative damage in vivo and in cell culture: how should you do it and what do the results mean? *Br J Pharmacol* 2004;142:231–255
23. Hozawa A, Jacobs DR Jr, Steffes MW, Gross MD, Steffen LM, Lee DH. Associations of serum carotenoid concentrations with the development of diabetes and with insulin concentration: interaction with smoking: the Coronary Artery Risk Development in Young Adults (CARDIA) Study. *Am J Epidemiol* 2006;163:929–937
24. Hozawa A, Jacobs DR Jr, Steffes MW, Gross MD, Steffen LM, Lee DH. Relationships of circulating carotenoid concentrations with several markers of inflammation, oxidative stress, and endothelial dysfunction: the Coronary Artery Risk Development in Young Adults (CARDIA)/Young Adult Longitudinal Trends in Antioxidants (YALTA) study. *Clin Chem* 2007;53:447–455
25. Gross M, Yu X, Hannan P, Prouty C, Jacobs DR Jr. Lipid standardization of serum fat-soluble antioxidant concentrations: the YALTA study. *Am J Clin Nutr* 2003;77:458–466