

Emerging Applications of Metabolomic and Genomic Profiling in Diabetic Clinical Medicine

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Clinical and epidemiological metabolomics provides a unique opportunity to look at genotype-phenotype relationships as well as the body's responses to environmental and lifestyle factors. Fundamentally, it provides information on the universal outcome of influencing factors on disease states and has great potential in the early diagnosis, therapy monitoring, and understanding of the pathogenesis of disease. Diseases, such as diabetes, with a complex set of interactions between genetic and environmental factors, produce changes in the body's biochemical profile, thereby providing potential markers for diagnosis and initiation of therapies. There is clearly a need to discover new ways to aid diagnosis and assessment of glycemic status to help reduce diabetes complications and improve the quality of life. Many factors, including peptides, proteins, metabolites, nucleic acids, and polymorphisms, have been proposed as putative biomarkers for diabetes. Metabolomics is an approach used to identify and assess metabolic characteristics, changes, and phenotypes in response to influencing factors, such as environment, diet, lifestyle, and pathophysiological states. The specificity and sensitivity using metabolomics to identify biomarkers of disease have become increasingly feasible because of advances in analytical and information technologies. Likewise, the emergence of high-throughput genotyping technologies and genome-wide association studies has prompted the search for genetic markers of diabetes predisposition or susceptibility. In this review, we consider the application of key metabolomic and genomic methodologies in diabetes and summarize the established, new, and emerging metabolomic and genomic biomarkers for the disease. We conclude by summarizing future insights into the search for improved biomarkers for diabetes research and human diagnostics.

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D iabetes is a rapidly increasing metabolic disorder precipitated by complex and poorly understood interactions between multiple environmental and genetic factors. The consequences of diabetes are far reaching, and disturbances in both the secretion and action of insulin impact on the global regulation of metabolism, affecting the composition of blood and other body fluids. Understanding of this process and identification of potential disease biomarkers have been greatly facilitated in recent years by the upsurge in new technologies for comprehensive metabolic profiling, which are often collectively termed metabolomics.

Metabolomic profiling in clinical medicine

Metabolomics is defined as the analytical description of biological samples

accompanied by the characterization and quantification of small molecules. It can often be confused with the term metabonomics, which represents the global, dynamic metabolic response of living systems to biological stimuli or genetic manipulation. Both terms are closely affiliated with each other owing to the analytical and experimental technologies used in each field. The observation of the characteristics and changes in metabolism by metabolomics allow the resulting data to be merged with data from the other “-omic” technologies. Genomic, metabolomic, and proteomic state-of-the-art technologies are now used increasingly by researchers to identify clinical methodologies for the early diagnosis and monitoring of human degenerative diseases such as diabetes. Classical risk factors still have an important role to play in diabetes assessment;

however, powerful methodologies are now available for exploitation of novel quantitative and qualitative disease-related biomarkers. Novel biomarkers are needed that are independent of known clinical risk factors.

Fundamentally, metabolomics aims to monitor changes in products of metabolism and provide valuable information on a range of influencing factors and gene-related outcomes. Exploitation of genomic technology in recent times has resulted in many technical advances, and genomic analysis has now emerged as a valuable tool in predicting the body's response to stimuli caused by disease or injury. Indeed, methodologies such as epigenetic profiling, sequencing technologies, microarrays, functional fingerprinting, and analysis of genomic alterations are all well-established methodologies in practice. Complementing these technologies with computational methods/bioinformatics that integrate large amounts of heterogeneous genetic and genomic information has helped provide meaningful results to aid our understanding of the complex changes of genes and macromolecules. There is now a clear need to discover novel and effective clinical biomarkers using technologies that encompass an array of different methodologies. Chromatography, two-dimensional electrophoresis, mass spectrometry, functional magnetic resonance, positron emission tomography, and protein/gene sequencing are some examples being used to unravel the body's complex biological systems. Sensitive and high-resolution techniques used in clinical metabolomics, such as nuclear magnetic resonance, gas chromatography–mass spectrometry, and liquid chromatography–mass spectrometry, are sensitive and robust and have the capacity to process large volumes of data from population studies (1,2). However, overinterpretation of data remains one of the key limitations to be overcome for successful exploitation of metabolomics and metabonomics.

In this brief review, we consider recent applications of metabolomic and related technologies in diabetes together with their use in relation to clinical diagnostics. Technical details of the methodologies involved and their use in basic diabetes research have

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Table 1—Established, new, and emerging metabolomic biomarkers for type 2 diabetes

Predictor	Abbreviation	Reference no.
Metabolic markers		
Insulin		7 and 8
Glucose		7 and 8
γ -Glutamyl transferase	GGT	4 and 8
Alanine aminotransferase	ALT	5 and 6
Ferritin	FTH1	7 and 8
Pancreatic polypeptide	PP	9
Fibronectin		10
Fetuin A		11
Sex hormone-binding globulin	SHBG	7 and 12
Free testosterone		12
Insulin-like growth factor I	IGF-I	13
Insulin receptor		8
Creatine kinase-MB	CKMB	8
MR-Pro atrial natriuretic peptide	MR_PRO_ANP	8
NT-Pro B-type natriuretic peptide	NT_PRO_BNP	8
B-type natriuretic peptide	BNP	8
Biomarkers of glycemia		
Glycated hemoglobin	HbA _{1c}	
Fructosamine		14
1,5-Anhydroglucitol	1,5AG	15
Glycated albumin		16
Glycated insulin		17
Glycosylated amylin		18
Glycated LDL		19
Markers of oxidative stress and nutrient status		
Glutathione	GSH	20
Advanced glycosylated end products receptor	RAGE	24
Ascorbic acid	Vitamin C	21
25-Hydroxyvitamin D	Vitamin D	22
Homocysteine		8
Branched-chain and aromatic amino acids	Leu, Ile, Val, Tyr, Phe	23
Lipid-related markers		
Leptin	LEP	6 and 25
Adiponectin	ADIPOQ	6, 8, and 25
Apolipoprotein B	ApoB	8
Apolipoprotein A	ApoA	8
Endothelial and inflammatory markers		
C-reactive protein	CRP	6–8 and 26
Interleukin-18	IL-18	8 and 27
Interleukin-1 receptor antagonist	IL-1ra	28
Interleukin-2 receptor antagonist	IL-2ra	7
Interleukin-6	IL-6	6–8
Plasminogen activator inhibitor-1	PAI-1	6 and 29
Cell adhesion molecule	CAM	30
Tissue plasminogen activator antigen	t-PA antigen, PLAT	31
Neopterin		8
Von Willebrand factor	vWF	31

been covered in several excellent articles and reviews (1,3).

Metabolomics applied to the clinical diagnosis and prognosis of diabetes

The American Diabetes Association officially recommends HbA_{1c} testing for the diagnosis and monitoring of diabetes, and

the global comparison of HbA_{1c} values is now possible as a result of the International Federation of Clinical Chemistry and Laboratory Medicine establishing true international reference methods for HbA_{1c} (in millimoles per mole) and the successful preparation of pure HbA_{1c} calibration material. However, there is clearly a need to

discover new markers, as illustrated in gestational diabetes mellitus, where there is a drive to reconsider diagnostic criteria recognizing the possibility of adverse pregnancy outcomes of milder levels of glucose intolerance than hitherto appreciated.

Metabolic markers of type 2 diabetes.

Whereas glucose and insulin are the most well-established biomarkers, there are many new and emerging biomarkers of diabetes (Table 1). Positive associations have been reported between serum γ -glutamyl transferase and incident type 2 diabetes (4). Pathophysiological mechanisms underlying how serum γ -glutamyl transferase relates to type 2 diabetes risk have not been elucidated, but insulin resistance, oxidative stress, and chronic low-grade systemic inflammation may be involved. Alanine aminotransferase (ALT) is elevated in some patients with type 2 diabetes independently of confounding factors such as obesity (5,6), and evidence now indicates that markers associated with fatty liver may predict future development of type 2 diabetes. In most cases of nonalcoholic fatty liver disease, the hepatic component of metabolic syndrome, ALT is elevated, and studies have associated raised ALT with metabolic syndrome and type 2 diabetes (5).

A strong association has been found between raised ferritin levels (below the range indicative of clinical hemochromatosis) and development of incident diabetes (7,8). Ferritin was associated with diabetes independently of established risk factors (age, BMI, sex, family history, physical inactivity, and smoking), as well as dietary factors and alcohol intake. The mechanism is thought to involve insulin resistance, free radical damage, and accumulation of iron in hepatocytes.

Pancreatic polypeptide, believed to act as a regulator of pancreatic and gastrointestinal functions, has been proposed as a possible marker of β -cell failure in diabetes (9). Likewise, fibronectin levels change in insulin resistance, and Amrein et al. (10) proposed that levels could be used in the diagnosis of insulin resistance and monitoring of disease progression. Fetuin-A, a hepatic secretory protein that binds the insulin receptor and inhibits insulin action, has been shown to be associated with incident diabetes independent of other markers of insulin resistance (11). Sex hormone-binding globulin (SHBG) is known to be downregulated by insulin, and low levels have been reported to reflect insulin resistance and incident diabetes in women (7,12). Population studies have shown that low testosterone levels are

commonly associated with the prediction of type 2 diabetes and the metabolic syndrome. Although the inverse association of testosterone with diabetes is partially mediated by SHBG, low testosterone is linked to diabetes via a bidirectional relationship with visceral fat, muscle, and possibly bone (12). IGF-I, which is involved in somatic growth, cellular differentiation, and regulation of metabolism, is potentially another marker of diabetes, given its insulin-like effects and involvement in glucose homeostasis. Large-scale gene association and prospective observational studies are needed to fully elucidate the involvement of IGF-I (13).

In a recent study of 31 novel biomarkers, Salomaa et al. (8) demonstrated an association between clinically incident diabetes and insulin receptor, creatine kinase-MB, MR-Pro atrial natriuretic peptide, NT-Pro B-type natriuretic peptide, and B-type natriuretic peptide. The utility of these as potential biomarkers and the nature of their links to diabetes clearly deserve further study.

Biomarkers of glycemia in diabetes. HbA_{1c} is the most widely known glycosylated protein in diabetes, and its assay has been the gold standard for the evaluation of glycemic status for many years. Limitations of the HbA_{1c} measurement do exist, but it remains an important tool in the management of diabetes along with self-monitored blood glucose profile data. Fructosamine is another marker used in practice (14) and refers to the ketoamine rearrangement product formed by the interaction of glucose with the ϵ -amino group on lysine residues of albumin. The assay is thought to be less accurate than HbA_{1c} because of factors affecting the half-life of its many components and is thus considered of less clinical value. Other markers of glycemic control that have been considered but not widely used are 1,5-anhydroglucitol (15) and glycated albumin (16).

Much interest has surrounded the role of glycosylated regulatory proteins as biomarkers. Because insulin glycation is dependent on the degree and duration of hyperglycemia, monitoring of glycemic status in diabetic patients using glycated insulin could aid approaches to the diagnosis, management, and treatment of diabetes (17). Other peptides such as amylin and amylin-like peptides have been disclosed as potentially useful in the detection and/or evaluation of diabetes. Glycosylated amylin (18) has been proposed as a predictor of the onset of diabetes in patients who otherwise show normal glycemic control, and another group has filed a patent based on

monoclonal antibodies against glycated LDL for monitoring glycemic control (19). **Markers of oxidative stress and nutrient status in type 2 diabetes.** Since diabetes is associated with overproduction of different reactive oxygen species leading to long-term development of diabetes complications, a number of candidate biomarkers have emerged (Table 1). Reduced levels of antioxidants such as glutathione, vitamin C, and vitamin E (20–22) and changes in serum malondialdehyde and activities of superoxide dismutase and glutathione peroxidase have been found in diabetic patients as well as changes in other oxidative stress biomarkers such as catalase, glutathione reductase, lipid peroxidation, and nitrite concentration (20).

Ascorbic acid (vitamin C) (21) and 25-hydroxyvitamin D (vitamin D) (22) are both associated with diabetes risk, but because of the many confounding determinants, levels need further investigation. Homocysteine also has promising links to diabetes (8). These and other biomarkers in Table 1 have yielded promising results, but most have been tested one at a time, with lack of independent validations. Many of these apparently “independent” risk factors may in fact be related by virtue of their common origins or shared metabolic pathways (6). There may even be different patterns of biomarkers of diabetes associated with early or late-stage diabetes. Recently, amino acid profiles were proposed as important in assessing diabetes risk as elevated levels of five amino acids were shown to predict the development of diabetes at early stages (23). Combinations of the five branched-chain and aromatic amino acids—leucine, isoleucine, valine, tyrosine, and phenylalanine—rather than a single amino acid, served as a more accurate predictor of diabetes risk (23).

In diabetes, advanced glycation end products form as a consequence of long-term hyperglycemia, and a number of truncated forms of the advanced glycation end product receptor (RAGE) have been identified. The C-terminally truncated form, named endogenous secretory RAGE, has potential as a biomarker and in the estimation of the risk of atherosclerotic disorders and occurrence of metabolic syndrome (24).

Lipid-related markers of type 2 diabetes. Adipokines are involved in a broad range of physiological processes such as insulin sensitivity, lipid metabolism, vascular hemostasis, blood pressure regulation, angiogenesis, and appetite control. Leptin and adiponectin are associated with increased

risk of type 2 diabetes even after adjustment for BMI, lifestyle factors, and cardiovascular disease (6,25). It is well recognized that adiponectin increases insulin sensitivity, regulates glucose and lipid metabolism, and enhances insulin action in the liver. Serum levels of adiponectin have been shown to decrease with increasing obesity, and interestingly, elevated adiponectin has been associated with a lower incidence of diabetes (6,25). Leptin is already a marker of percentage fat mass in healthy individuals and regulates body weight by effects on food intake and metabolism. The association between leptin and incident diabetes has been difficult to determine but may also reflect insulin resistance. Adiponectin is more strongly associated with type 2 diabetes risk than leptin (25). Apolipoprotein (Apo)B, and to a lesser extent ApoA, was a particularly strong predictor of diabetes even when controlling for BMI and waist-to-hip ratio (8).

Endothelial and inflammatory markers of type 2 diabetes. C-reactive protein (CRP) is a predictor of diabetes independent of other clinical indicators such as BMI, fasting triglyceride, and glucose (26), but circulating levels correlate with lipids, SHBG, and adiponectin. The value of CRP is promising, but further evaluation is needed. Elevated levels of the cytokine interleukin (IL)-18 are linked with an increased risk of type 2 diabetes, independent of a generalized proinflammatory state (27). Studies have also reported upregulation of anti-inflammatory cytokine IL-1 receptor antagonist (IL-1ra) in individuals with obesity and insulin resistance (28). These studies indicate that individuals with high risk of type 2 diabetes can be characterized by the presence of an early compensatory, anti-inflammatory response that precedes the development of the disease and inflammatory markers. Like IL-1ra and CRP, IL-2ra is involved in inflammatory pathways; however, only one study to date has identified IL-2ra as a diabetes marker (7), which may be due to oxidative stress in diabetes culminating in T lymphocyte activation.

IL-6 has been reported to be elevated in incident diabetes, independent of obesity and fasting glucose (6–8). Studies are needed to determine the relationship between IL-6 and diabetes and whether there is a causal link. Plasminogen activator inhibitor 1 levels reflect an acute phase response, and elevated levels are found with incident diabetes independent of obesity and insulin resistance (29). The association between high plasminogen

activator inhibitor 1 and incident diabetes may be due to associations with liver fat (6). Circulating levels of several other inflammatory endothelial-derived factors such as cell adhesion molecules (30), tissue-plasminogen activator antigen (31), neopterin (8), and von Willebrand factor (31) have been linked to diabetes risk. Recent studies such as the MONICA/KORA study have shown that when a risk prediction model of multiple inflammation markers is used, the prediction of incident type 2 diabetes and coronary events is significantly improved compared with cardiometabolic risk factors (25,27).

Increased clinical value of evaluating panels composed of different biomarkers

Advances in technology plus awareness of an increasing number of diabetes-related metabolomic analytes are likely to facilitate use of panels of combined biomarkers rather than reliance on single biomarkers for diabetes. If these are selected from different tissue origins/pathways, their ability to predict diabetes risk is likely to be increased, thereby facilitating earlier interventions. This view is supported by two comprehensive studies (7,8). Kollberg et al. (7) evaluated the potential of 58 candidate diabetes-related biomarkers plus six clinical factors for predicting 5-year risk of diabetes in 160 of 632 individuals from the Danish Inter99 cohort who went on to develop type 2 diabetes. A six-biomarker model (adiponectin, CRP, ferritin, IL-2ra, glucose, and insulin) showed improved performance over single markers such as HbA_{1c} and fasting glucose, being equivalent to a 2-h oral glucose tolerance test (7). Similarly, Salomaa et al. (8) evaluated the potential of 31 novel biomarkers as predictors for clinically incident diabetes in a combined total of 12,804 individuals from the FINRISK97 and Health 2000 cohorts of whom 596 later developed diabetes in 10-year follow-up. This study revealed that adiponectin, ApoB, CRP, and ferritin improved diabetes prediction even after taking BMI, glucose, and other classical risk factors into account (8). Sex-specific analysis further showed potential value of including IL-1ra and insulin as biomarkers. These data suggest that biomarker scores reflecting different pathological processes may have significant potential for improving future prediction of diabetes. Similarly, evaluation of amino acid profiles appears to be more effective in prediction than are single amino acids (24).

Genomic variations and DNA profiling of those at risk for type 2 diabetes

Despite many candidate gene studies and genome-wide linkage studies, very few susceptibility loci for type 2 diabetes have been identified until the recent emergence of genomic-wide association (GWA) data and large-scale replication studies (Table 2). Meta-analysis of GWA studies provides the unique opportunity to investigate the heterogeneity or consistency of genomic associations across diverse datasets and study populations. Recently, Voight et al. (32), using large-scale association analyses combining the data from eight GWA studies, identified 12 new susceptibility loci for type 2 diabetes.

Despite identification of many putative causative genetic variants, few have generated credible susceptibility variants for type 2 diabetes. Indeed, the most important finding using linkage studies is the discovery that the alteration of *TCF7L2* (*TCF-4*) gene expression or function (33) disrupts pancreatic islet function and results in enhanced risk of type 2 diabetes. Candidate gene studies have also reported many type 2 diabetes-associated loci and the coding variants in the nuclear receptor peroxisome proliferator-activated receptor- γ (34), the potassium channel *KCNJ11* (34), *WFS1* (35), and *HNF1B* (*TCF2*) (36) are among the few that have been replicated (Table 2). Recently, there have been great advances in the analysis of associated variants in GWA and replication studies due to high-throughput genotyping technologies, the International HapMap Project, and the Human Genome Project. Type 2 susceptibility loci such as *JAZF1*, *CDC123-CAMK1D*, *TSPAN8-LGR5*, *THADA*, *ADAMTS9*, *NOTCH2*, and *ADCY5* (37,38) are among some of the established loci (Table 2). *CDKN2A/B*, *CDKAL1*, *SLC30A8*, *IGF2BP2*, *HHEX/IDE*, and *FTO* are other established susceptibility loci for diabetes (Table 2) (34,39,40). GWA studies have also identified the potassium voltage-gated channel *KCNQ1* (32) as an associated gene variant for diabetes. A recent GWA study reporting a genetic variant with a strong association with insulin resistance, hyperinsulinemia, and type 2 diabetes, located adjacent to the insulin receptor substrate 1 (*IRS1*) gene, is the C allele of rs2943641 (41). Interestingly, the parental origin of the single nucleotide polymorphism is of importance because the allele that confers risk when paternally inherited is protected when maternally transmitted. GWA studies for glycemic traits have identified loci such as *MTNR1B*

(42), *GCK* (glucokinase) (42), and *GCKR* (glucokinase receptor) (42); however, further investigation of genetic loci on glucose homeostasis and their impact on type 2 diabetes is needed. Indeed, a recent study by Soranzo et al. (42) using GWA studies identified ten genetic loci associated with HbA_{1c}. Genetic factors affecting expression, turnover, and abnormal glycation of hemoglobin may be associated with changes in levels of HbA_{1c}.

Significant effects of many susceptibility loci are still to be determined and replicated, and further large-scale association studies will be required. Recently, Schleinitz et al. (43) found some of the type 2 diabetes risk alleles or related subphenotypes to be weak, including those of *JAZF1*, *CDC123/CAMK1D*, *NOTCH2*, *ADAMTS9*, *THADA*, and *TSPAN8-LGR5*. The *TNF/LTA* locus has been a long-standing type 2 diabetes candidate gene, whereas a recent study found no evidence of an association between *TNF/LTA* region variation and type 2 diabetes (44). The association of polymorphisms in *TNFA* and type 2 diabetes has been extensively reported. Recently, the *TNFA* variant rs3093662, linked to higher serum levels of tumor necrosis factor- α , was shown to be associated with elevated insulin (45). Mutated transcription factors, hepatocyte nuclear factor (*HNF*)1A and *HNF4A*, have received substantial attention, and there is evidence for susceptibility of the variants to maturity-onset diabetes of the young (MODY) and type 2 diabetes. Recently, high-sensitivity CRP was shown to discriminate *HNF1A*-MODY from other subtypes of diabetes (46).

Interestingly, many of the established susceptibility loci are involved in insulin secretion signaling, supporting an important role for defects in β -cell function and β -cell mass in type 2 diabetes. The exciting potential of genetic testing for susceptibility of diabetes appears to be some way off, apart from rare forms of monogenic diabetes (44). Moreover, it is well known that non-genetic factors such as obesity and lifestyle factors play an important role in the disease. New phenotyping approaches to studying metabolite and protein abundance and data integration are needed to bring genomic and metabolomic goals together. In this context, the Human Metabolome Project in Canada (47), aimed at providing a linkage between the human metabolome and the human genome, has identified and quantified normal concentration ranges for a large number of metabolites in cerebrospinal fluid, serum, urine, and other

Table 2—Type 2 diabetes susceptibility loci established through candidate-gene, genome-wide linkage, and GWA studies

Gene/region	Gene name	Chromosomal location	Identification	Reference no.
<i>TCF7L2</i>	Transcription factor 7-like 2	10q25.3	Linkage study	33 and 39
<i>PPARγ</i>	Peroxisome proliferator-activated receptor γ	3q25	Candidate gene	34
<i>KCNJ11</i>	Potassium channel, inwardly rectifying subfamily J, member 11	11p15.5	Candidate gene	34
<i>WFS1</i>	Wolfram syndrome 1 (wolframin)	4p16.1	Candidate gene	35
<i>HNF1B</i>	HNF1 homeobox B	17q12	Candidate gene	36
<i>JAZF1</i>	Juxtaposed with another zinc finger gene 1	7p15	GWA	37
<i>CDC123-CAMK1D</i>	Cell division cycle protein 123 homolog/calcium/calmodulin-dependent protein kinase 1D	10p13-p14	GWA	37
<i>TSPAN8-LGR5</i>	Tetraspanin 8 and leucine-rich-repeat-containing G-protein coupled	12q21	GWA	37
<i>THADA</i>	Thyroid adenoma-associated	2p21	GWA	37
<i>ADAMS9</i>	ADAM metalloproteinase with thrombospondin type 1 motif, 9	3p14	GWA	37
<i>NOTCH2</i>	Notch homolog 2, <i>Drosophila</i>	1p12	GWA	37
<i>ADCY5</i>	Adenylate cyclase	3	GWA	38
<i>CDKN2A/B</i>	Cyclin-dependent kinase inhibitor 2A/B	9p21	GWA	40
<i>CDKAL1</i>	CDK5 regulatory subunit associated protein 1-like 1	6p22.2	GWA	34 and 40
<i>SLC30A8</i>	Solute carrier family 30, member 8	8q24.11	GWA	34 and 40
<i>IGF2BP2</i>	Insulin-like growth factor 2 mRNA binding protein 2	3q28	GWA	34 and 40
<i>HHEX/IDE</i>	Hematopoietically expressed homeobox and insulin-degrading enzyme	10q23-q25	GWA	34 and 40
<i>FTO</i>	Fat mass and obesity associated	16q12.2	GWA	34
<i>MTNR1B</i>	Melatonin receptor 1B	11q21-q22	GWA	42
<i>KCNQ1</i>	Potassium channel, voltage-gated, KQT-like subfamily, member 1	12q21	GWA	32
<i>IRS1</i>	Insulin receptor substrate 1	2q36	GWA	41
<i>GCK</i>	Glucokinase	7p15.3-p15.1	GWA	42
<i>GCKR</i>	Glucokinase regulator	2p23	GWA	42
<i>C6PC2</i>	Glucose-6-phosphatase, catalytic 2	2q24.3	GWA	42
<i>TNFA</i>	Tumor necrosis factor- α	6p21.3	Candidate gene	45
<i>HNF1A</i>	Hepatocyte nuclear factor 1 α	12q24.2	Candidate gene	46
<i>HNF4A</i>	Hepatocyte nuclear factor 4 α	20q13.12	Candidate gene	46

tissues and biofluids. There are currently 7,900 entries in the Human Metabolome Database (<http://www.hmdb.ca>), which contains detailed information about small molecule metabolites and will be useful for applications in metabolomics, clinical chemistry, and biomarker discovery.

Susceptibility gene markers in type 1 diabetes

In type 1 diabetes, the study of susceptibility genes has been facilitated by the availability of large collections of families with affected sibling pairs as seen in the Type 1 Diabetes Genetics Consortium (48). Type 1 diabetes is a multifactorial disease where loci within the HLA account for most of the genetic susceptibility. The major susceptibility locus maps to the HLA class II genes at 6p21, accounting

for up to 30–50% of genetic type 1 diabetes risk (48). The association of genes of the class II region is thought to reflect their role in the T-cell immune response. The major susceptibility class II loci are HLA-DRB1 and HLA-DQB1/DQA1 on chromosome 6p21 and, to a lesser extent, HLA-DPB1/DPA1 (48,49). Association studies are complicated by the high polymorphism of the HLA DPA1 and DPB1 loci. The HLA loci, DRB1, DQA1, DQB1, DPA1, DPB1, A, B, and C, have repeatedly been shown to be involved in type 1 diabetes susceptibility. However, conflicting results have been obtained for the HLA loci involved in susceptibility or protection as a result of coinherited loci, population-specific differences, typing approaches used, differences in study design, or low-powered studies (49).

The highest-risk DR/DQ haplotypes, DR3 and DR4, exhibit a spectrum of risk from increased to neutral to protective (50). Type 1 diabetes incidence is increasing worldwide each year, and it appears that as the disease increases the percentage of cases with the high-risk HLA DR3/4 genotype is decreasing, suggesting an increased environmental pressure or contribution of other non-HLA class II alleles to diabetes risk (51). Type 1 diabetes risk is also linked to the major histocompatibility complex independently of HLA-DR/DQ such as HLA class I alleles (52). Studies have supported a role for HLA class I alleles in type 1 diabetes susceptibility including B*3906 and B*5701 (53). The large datasets generated by studies such as the Type 1 Diabetes Genetics Consortium are crucial for the generation of sufficient class I data

for disease association studies (48). Studies are ongoing, investigating HLA class I and class III alleles.

More than 40 non-HLA susceptibility gene markers have been identified that contribute to type 1 diabetes risk. However, for many of these genetic predictors of risk the effect is small, even for the strongest loci (54). These non-MHC loci include the insulin gene (*INS*) on chromosome 11p15 (55), which confers ~10% of the genetic susceptibility to type 1 diabetes. The cytotoxic T cell-associated protein 4 (*CTLA4*) gene on chromosome 2q33 is associated with type 1 diabetes (56). Also, the protein tyrosine phosphatase, nonreceptor type 22 (lymphoid) (*PTPN22*) gene on chromosome 1p13 (57), involved in preventing spontaneous T-cell activation, is linked to type 1 diabetes risk (57). A number of other associations have been proposed such as IL-2 receptor, α (*IL2RA*), and interferon-induced with helicase C domain 1 (*IFIH1*) genes (58) and KIAA0350 (59) and small ubiquitin-like modifier 4 (*SUMO4*) (60). Many new candidate genes are emerging such as *IL10*, *IL19*, *IL20*, *GLIS3*, *CD69*, and *IL27* (58), but further genotyping and functional studies are needed to determine whether the genes are causal. With important breakthroughs in DNA sequencing technology and mapping of diabetes cases, the determination of extreme genetic risk of type 1 diabetes in the general population could eventually lead to intervention or prevention trials.

Future developments in the search for improved biomarkers for the diagnosis and treatment of diabetes

Translating research findings to useful and reliable clinical tests has been challenging; however, the discovery of ideal biomarkers for diabetes is improving along with the development of biomarker panels and new methodologies. In the future, diagnostic tests may be used to select individuals who are likely to benefit from treatment or those who demonstrate an objective indication of treatment efficacy. Emergence of diabetes-associated genetic variants represents a powerful tool for improving our understanding of the pathogenesis of diabetes. However, translation of these novel findings to genetic screening and personalized medicine is still at an early stage. Characterization of functional variants and an understanding of the mechanisms by which these loci confer susceptibility to disease are needed. With discovery of genes linked to fasting glucose, it may be possible to identify other loci

associated with additional features of the type 2 diabetes phenotype such as impaired glucose tolerance, defective first-phase insulin release, and insulin resistance. Combination of genetic and biomarkers screens may provide further opportunities. The challenges in harnessing the potential of new biomarkers should be alleviated by new and exciting collaborations between pharmaceutical agencies, diagnostic companies, and academic institutions, with the harnessing of skills from the different clinical, biomedical, diagnostic, and pharmacological areas.

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References

1. Lanza IR, Zhang S, Ward LE, Karakelides H, Raftery D, Nair KS. Quantitative metabolomics by H-NMR and LC-MS/MS confirms altered metabolic pathways in diabetes. *PLoS ONE* 2010;5:e10538
2. Li LO, Hu YF, Wang L, Mitchell M, Berger A, Coleman RA. Early hepatic insulin resistance in mice: a metabolomics analysis. *Mol Endocrinol* 2010;24:657–666
3. Bain JR, Stevens RD, Wenner BR, Ilkayeva O, Muoio DM, Newgard CB. Metabolomics applied to diabetes research: moving from information to knowledge. *Diabetes* 2009;58:2429–2443
4. Meisinger C, Löwel H, Heier M, Schneider A, Thorand B; KORA Study Group. Serum gamma-glutamyltransferase and risk of type 2 diabetes mellitus in men and women from the general population. *J Intern Med* 2005;258:527–535
5. Schindhelm RK, Diamant M, Dekker JM, Tushuizen ME, Teerlink T, Heine RJ. Alanine aminotransferase as a marker of non-alcoholic fatty liver disease in relation to type 2 diabetes mellitus and cardiovascular disease. *Diabetes Metab Res Rev* 2006;22:437–443
6. Sattar N, Wannamethee SG, Forouhi NG. Novel biochemical risk factors for type 2 diabetes: pathogenic insights or prediction possibilities? *Diabetologia* 2008;51:926–940
7. Kolberg JA, Jørgensen T, Gerwien RW, et al. Development of a type 2 diabetes risk

model from a panel of serum biomarkers from the Inter99 cohort. *Diabetes Care* 2009;32:1207–1212

8. Salomaa V, Havulinna A, Saarela O, et al. Thirty-one novel biomarkers as predictors for clinically incident diabetes. *PLoS ONE* 2010;5:e10100
9. Christ A, Evers S, Krapfenbauer K, Sebokova E. Pancreatic polypeptide as target/marker of beta cell failure. US patent 2009/0215069. 27 August 2009
10. Amrein K, Berndt P, Evers S, Foster S, Fountoulakis M, Sebokova E. Fibronectin as target/marker for insulin resistance. International patent WO/2007/06278. 7 June 2007
11. Stefan N, Fritsche A, Weikert C, et al. Plasma fetuin-A levels and the risk of type 2 diabetes. *Diabetes* 2008;57:2762–2767
12. Ding EL, Song Y, Malik VS, Liu S. Sex differences of endogenous sex hormones and risk of type 2 diabetes: a systematic review and meta-analysis. *JAMA* 2006;295:1288–1299
13. Sandhu MS. Insulin-like growth factor-I and risk of type 2 diabetes and coronary heart disease: molecular epidemiology. *Endocr Dev* 2005;9:44–54
14. Armbruster DA. Fructosamine: structure, analysis, and clinical usefulness. *Clin Chem* 1987;33:2153–2163
15. Nowatzke W, Samo MJ, Birch NC, Stickle DF, Eden T, Cole TG. Evaluation of an assay for serum 1,5-anhydroglucitol (GlycoMark) and determination of reference intervals on the Hitachi 917 analyzer. *Clin Chim Acta* 2004;350:201–209
16. Kouzuma T, Usami T, Yamakoshi M, Takahashi M, Imamura S. An enzymatic method for the measurement of glycated albumin in biological samples. *Clin Chim Acta* 2002;324:61–71
17. Lindsay JR, McKillop AM, Mooney MH, O'Harte FPM, Bell PM, Flatt PR. Demonstration of increased concentrations of circulating glycated insulin in human type 2 diabetes using a novel and specific radioimmunoassay. *Diabetologia* 2003;46:475–478
18. Rittenhouse J, Koda J, Fineman M, Percy A. Glycosylated amylin. International patent WO/1997/035600. 2 October 1997
19. Cohen MP. Monoclonal antibodies against glycated low density lipoprotein. US patent 5,494,791. 27 February 1996
20. Maritim AC, Sanders RA, Watkins JB 3rd. Diabetes, oxidative stress, and antioxidants: a review. *J Biochem Mol Toxicol* 2003;17:24–38
21. Sargeant LA, Wareham NJ, Bingham S, et al. Vitamin C and hyperglycemia in the European Prospective Investigation into Cancer—Norfolk (EPIC-Norfolk) study: a population-based study. *Diabetes Care* 2000;23:726–732
22. Zhao G, Ford ES, Li C. Associations of serum concentrations of 25-hydroxyvitamin D and parathyroid hormone with surrogate markers of insulin resistance among U.S.

- adults without physician-diagnosed diabetes: NHANES, 2003-2006. *Diabetes Care* 2010;33:344-347
23. Wang TJ, Larson MG, Vasan RS, et al. Metabolite profiles and the risk of developing diabetes. *Nat Med* 2011;17:448-453
 24. Koyama H, Yamamoto H, Nishizawa Y. Endogenous secretory RAGE as a novel biomarker for metabolic syndrome and cardiovascular diseases. *Biomark Insights* 2007;2:331-339
 25. Thorand B, Zierer A, Baumert J, Meisinger C, Herder C, Koenig W. Associations between leptin and the leptin/adiponectin ratio and incident type 2 diabetes in middle-aged men and women: results from the MONICA/KORA Augsburg study 1984-2002. *Diabet Med* 2010;27:1004-1011
 26. Freeman DJ, Norrie J, Caslake MJ, et al.; West of Scotland Coronary Prevention Study. C-reactive protein is an independent predictor of risk for the development of diabetes in the West of Scotland Coronary Prevention Study. *Diabetes* 2002;51:1596-1600
 27. Thorand B, Kolb H, Baumert J, et al. Elevated levels of interleukin-18 predict the development of type 2 diabetes: results from the MONICA/KORA Augsburg Study, 1984-2002. *Diabetes* 2005;54:2932-2938
 28. Herder C, Brunner EJ, Rathmann W, et al. Elevated levels of the anti-inflammatory interleukin-1 receptor antagonist precede the onset of type 2 diabetes: the Whitehall II study. *Diabetes Care* 2009;32:421-423
 29. Festa A, D'Agostino R Jr, Tracy RP, Haffner SM; Insulin Resistance Atherosclerosis Study. Elevated levels of acute-phase proteins and plasminogen activator inhibitor-1 predict the development of type 2 diabetes: the insulin resistance atherosclerosis study. *Diabetes* 2002;51:1131-1137
 30. Song Y, Manson JE, Tinker L, et al. Circulating levels of endothelial adhesion molecules and risk of diabetes in an ethnically diverse cohort of women. *Diabetes* 2007;56:1898-1904
 31. Wannamethee SG, Sattar N, Rumley A, Whincup PH, Lennon L, Lowe GD. Tissue plasminogen activator, von Willebrand factor, and risk of type 2 diabetes in older men. *Diabetes Care* 2008;31:995-1000
 32. Voight BF, Scott LJ, Steinthorsdottir V, et al.; MAGIC investigators; GIANT Consortium. Twelve type 2 diabetes susceptibility loci identified through large-scale association analysis. *Nat Genet* 2010;42:579-589
 33. Grant SF, Thorleifsson G, Reynisdottir I, et al. Variant of transcription factor 7-like 2 (TCF7L2) gene confers risk of type 2 diabetes. *Nat Genet* 2006;38:320-323
 34. Scott LJ, Mohlke KL, Bonnycastle LL, et al. A genome-wide association study of type 2 diabetes in Finns detects multiple susceptibility variants. *Science* 2007;316:1341-1345
 35. Sandhu MS, Weedon MN, Fawcett KA, et al. Common variants in WFS1 confer risk of type 2 diabetes. *Nat Genet* 2007;39:951-953
 36. Winckler W, Weedon MN, Graham RR, et al. Evaluation of common variants in the six known maturity-onset diabetes of the young (MODY) genes for association with type 2 diabetes. *Diabetes* 2007;56:685-693
 37. Zeggini E, Scott LJ, Saxena R, et al.; Wellcome Trust Case Control Consortium. Meta-analysis of genome-wide association data and large-scale replication identifies additional susceptibility loci for type 2 diabetes. *Nat Genet* 2008;40:638-645
 38. Saxena R, Hivert MF, Langenberg C, et al.; GIANT consortium; MAGIC investigators. Genetic variation in GIPR influences the glucose and insulin responses to an oral glucose challenge. *Nat Genet* 2010;42:142-148
 39. Salonen JT, Uimari P, Aalto JM, et al. Type 2 diabetes whole-genome association study in four populations: the DiaGen consortium. *Am J Hum Genet* 2007;81:338-345
 40. Zeggini E, Weedon MN, Lindgren CM, et al.; Wellcome Trust Case Control Consortium (WTCCC). Replication of genome-wide association signals in UK samples reveals risk loci for type 2 diabetes. *Science* 2007;316:1336-1341
 41. Rung J, Cauchi S, Albrechtsen A, et al. Genetic variant near IRS1 is associated with type 2 diabetes, insulin resistance and hyperinsulinemia. *Nat Genet* 2009;41:1110-1115
 42. Soranzo N, Sanna S, Wheeler E, et al.; WTCCC. Common variants at 10 genomic loci influence hemoglobin A_{1c} levels via glycemic and nonglycemic pathways. *Diabetes* 2010;59:3229-3239
 43. Schleinitz D, Tönjes A, Böttcher Y, et al. Lack of significant effects of the type 2 diabetes susceptibility loci JAZF1, CDC123/CAMK1D, NOTCH2, ADAMTS9, THADA, and TSPAN8/LGR5 on diabetes and quantitative metabolic traits. *Horm Metab Res* 2010;42:14-22
 44. Boraska V, Rayner NW, Groves CJ, et al. Large-scale association analysis of TNF/LTA gene region polymorphisms in type 2 diabetes. *BMC Med Genet* 2010;11:69
 45. Arora P, Garcia-Bailo B, Dastani Z, et al. Genetic polymorphisms of innate immunity-related inflammatory pathways and their association with factors related to type 2 diabetes. *BMC Med Genet* 2011;12:95
 46. McDonald TJ, Shields BM, Lawry J, Owen KR, Gloyn AL, Ellard S, Hattersley AT. High-sensitivity CRP discriminates HNF1A-MODY from other subtypes of diabetes. *Diabetes Care* 2011;34:1860-1862
 47. Psychogios N, Hau DD, Peng J, et al. The human serum metabolome. *PLoS ONE* 2011;6:e16957
 48. Erlich H, Valdes AM, Noble J, et al.; Type 1 Diabetes Genetics Consortium. HLA DR-DQ haplotypes and genotypes and type 1 diabetes risk: analysis of the type 1 diabetes genetics consortium families. *Diabetes* 2008;57:1084-1092
 49. Varney MD, Valdes AM, Carlson JA, et al.; Type 1 Diabetes Genetics Consortium. HLA DPA1, DPB1 alleles and haplotypes contribute to the risk associated with type 1 diabetes: analysis of the type 1 diabetes genetics consortium families. *Diabetes* 2010;59:2055-2062
 50. Pugliese A, Gianani R, Moromisato R, et al. HLA-DQB1*0602 is associated with dominant protection from diabetes even among islet cell antibody-positive first-degree relatives of patients with IDDM. *Diabetes* 1995;44:608-613
 51. Steck AK, Armstrong TK, Babu SR, Eisenbarth GS; Type 1 Diabetes Genetics Consortium. Stepwise or linear decrease in penetrance of type 1 diabetes with lower-risk HLA genotypes over the past 40 years. *Diabetes* 2011;60:1045-1049
 52. Noble JA, Martin A, Valdes AM, et al. Type 1 diabetes risk for human leukocyte antigen (HLA)-DR3 haplotypes depends on genotypic context: association of DPB1 and HLA class I loci among DR3- and DR4-matched Italian patients and controls. *Hum Immunol* 2008;69:291-300
 53. Valdes AM, Erlich HA, Noble JA. Human leukocyte antigen class I B and C loci contribute to Type 1 Diabetes (T1D) susceptibility and age at T1D onset. *Hum Immunol* 2005;66:301-313
 54. Steck AK, Rewers MJ. Genetics of type 1 diabetes. *Clin Chem* 2011;57:176-185
 55. Bell GI, Horita S, Karam JH. A polymorphic locus near the human insulin gene is associated with insulin-dependent diabetes mellitus. *Diabetes* 1984;33:176-183
 56. Concannon P, Chen WM, Julier C, et al.; Type 1 Diabetes Genetics Consortium. Genome-wide scan for linkage to type 1 diabetes in 2,496 multiplex families from the Type 1 Diabetes Genetics Consortium. *Diabetes* 2009;58:1018-1022
 57. Bottini N, Musumeci L, Alonso A, et al. A functional variant of lymphoid tyrosine phosphatase is associated with type 1 diabetes. *Nat Genet* 2004;36:337-338
 58. Barrett JC, Clayton DG, Concannon P, et al. Genome-wide association study and meta-analysis find that over 40 loci affect risk of type 1 diabetes. *Nat Genet* 2009;41:703-707
 59. Concannon P, Onengut-Gumuscu S, Todd JA, et al.; Type 1 Diabetes Genetics Consortium. A human type 1 diabetes susceptibility locus maps to chromosome 21q22.3. *Diabetes* 2008;57:2858-2861
 60. Guo D, Li M, Zhang Y, et al. A functional variant of SUMO4, a new I kappa B alpha modifier, is associated with type 1 diabetes. *Nat Genet* 2004;36:837-841