Diabetes Care





Pioglitazone Improves Left Ventricular Diastolic Function in Subjects With Diabetes

https://doi.org/10.2337/dc17-0078

Geoffrey D. Clarke,^{1,2}
Carolina Solis-Herrera,¹
Marjorie Molina-Wilkins,¹
Sandra Martinez,¹ Aurora Merovci,¹
Eugenio Cersosimo,¹ Robert J. Chilton,^{3,4}
Patricia Iozzo,⁵ Amalia Gastaldelli,^{1,5}
Muhammad Abdul-Ghani,¹ and
Ralph A. DeFronzo^{1,4}

OBJECTIVE

To examine the effect of pioglitazone on myocardial insulin sensitivity and left ventricular (LV) function in patients with type 2 diabetes (T2D).

RESEARCH DESIGN AND METHODS

Twelve subjects with T2D and 12 with normal glucose tolerance received a euglycemic insulin clamp. Myocardial glucose uptake (MGU) and myocardial perfusion were measured with [¹⁸F]fluoro-2-deoxy-D-glucose and [¹⁵O]H₂O positron emission tomography before and after 24 weeks of pioglitazone treatment. Myocardial function and transmitral early diastolic relation/atrial contraction (E/A) flow ratio were measured with magnetic resonance imaging.

RESULTS

Pioglitazone reduced HbA $_{1c}$ by 0.9%; decreased systolic and diastolic blood pressure by 7 \pm 2 and 7 \pm 2 mmHg, respectively (P < 0.05); and increased whole-body insulinstimulated glucose uptake by 71% (3.4 \pm 1.3 to 5.8 \pm 2.1 mg/kg \cdot min; P < 0.01) in subjects with T2D. Pioglitazone enhanced MGU by 75% (0.24 \pm 0.14 to 0.42 \pm 0.13 μ mol/min \cdot g; P < 0.01) and myocardial perfusion by 16% (0.95 \pm 0.16 to 1.10 \pm 0.25 mL/min \cdot g; P < 0.05). Measures of diastolic function, E/A ratio (1.04 \pm 0.3 to 1.25 \pm 0.4) and peak LV filling rate (349 \pm 107 to 433 \pm 99 mL/min), both increased (P < 0.01). End-systolic volume, end-diastolic volume, peak LV ejection rate, and cardiac output trended to increase (P not significant), whereas the ejection fraction (61 \pm 6 to 66 \pm 7%) and stroke volume increased significantly (71 \pm 20 to 80 \pm 20 L/min; both P < 0.05).

CONCLUSIONS

Pioglitazone improves whole-body and myocardial insulin sensitivity, LV diastolic function, and systolic function in T2D. Improved myocardial insulin sensitivity and diastolic function are strongly correlated.

The incidence of cardiovascular (CV) disease, including myocardial infarction, stroke, and heart failure, is increased two- to threefold in patients with type 2 diabetes (T2D) (1,2). Heart failure is an ominous sign in patients with diabetes, and 50% of those with T2D and heart failure die within 5 years (3). Diastolic dysfunction is a common abnormality in T2D found on echocardiography, yet most patients with T2D are asymptomatic (4–6).

Peripheral tissues, including skeletal (7) and cardiac (8,9) muscle, liver (10), and adipose tissue (11), are resistant to insulin in T2D. We and others have shown that pioglitazone is a potent insulin sensitizer in skeletal muscle, liver, and adipocytes (7,12–16). However, only one study examined the effect of pioglitazone on myocardial

Received 11 January 2017 and accepted 22 July 2017.

Clinical trial reg. no. NCT0588470, clinicaltrials

This article contains Supplementary Data online at http://care.diabetesjournals.org/lookup/suppl/doi:10.2337/dc17-0078/-/DC1.

G.D.C. and C.S.-H. contributed equally to this work.

© 2017 by the American Diabetes Association. Readers may use this article as long as the work is properly cited, the use is educational and not for profit, and the work is not altered. More information is available at http://www.diabetesjournals.org/content/license.

¹Diabetes Division, University of Texas Health Science Center, San Antonio, TX

²Department of Radiology, University of Texas Health Science Center, San Antonio, TX

³Division of Cardiology, University of Texas Health Science Center, San Antonio, TX

⁴South Texas Veterans Health Care System, San Antonio, TX

⁵Institute of Clinical Physiology, Pisa, Italy

Corresponding author: Ralph A. DeFronzo, albarado@uthscsa.edu.

insulin sensitivity and demonstrated that pioglitazone increases myocardial insulin sensitivity and parameters of cardiac function, but no correlation between improved cardiac function and increase in myocardial insulin sensitivity was observed (8). The effect of pioglitazone on myocardial insulin sensitivity in T2D is of considerable clinical importance because myocardial insulin resistance has been implicated in the development of myocardial dysfunction and accelerated coronary atherosclerosis (4,5,17–19).

In the Prospective Pioglitazone Clinical Trial in Macrovascular Events (PROactive), pioglitazone significantly reduced the main second end point (CV death, nonfatal myocardial infarction, nonfatal stroke) (hazard ratio 0.84; P = 0.017), although the decrease in primary end point (major adverse CV events plus peripheral vascular disease) did not reach statistical significance because of an increase in leg revascularization (20), which we now know is refractory to glucose-, lipid-, and blood pressure-lowering therapies (21-23). In PROactive, the incidence of heart failure in pioglitazone-treated subjects was stated to be increased (20), but heart failure was not adjudicated, and overall mortality and CV events tended to decline (not increase) in the pioglitazonetreated group with heart failure. Because heart failure is an ominous sign in T2D, with 5-year mortality of \sim 50% (24), that these subjects really had heart failure is unlikely. The recently published Insulin Resistance Intervention After Stroke trial results are consistent with those of PROactive (25). In 3,876 patients with recent stroke or transient ischemic attack, pioglitazone reduced the incidence of recurrent stroke and cardiovascular events by 24% (P < 0.001). No difference in incidence of heart failure (P = 0.80) or hospitalization for heart failure (P = 0.35) was observed.

In the current study, we used cardiac magnetic resonance imaging (MRI) to quantify LV diastolic and systolic function and the euglycemic insulin clamp with positron emission tomography (PET) to quantify whole-body (primarily reflects muscle) and myocardial insulin sensitivity before and after pioglitazone treatment. Contrary to common belief (13,26-29), we hypothesized that pioglitazone improves—not impairs—parameters of LV diastolic and systolic function.

RESEARCH DESIGN AND METHODS Subjects

Twelve patients with T2D (age 51 \pm 9 years, 10 males, 2 females, HbA_{1c} $6.8 \pm 1.6\%$, diabetes duration 4.0 ± 3.1 years, BMI 30.8 \pm 4.3 kg/m², 7 Mexican American, 5 Caucasian) without clinically manifested CV disease participated in the study (Table 1). Subjects with T2D were drug naïve (n = 4) or treated with metformin (n = 7) or metformin/sulfonylurea (n = 1). We did not observe any differences regarding the effect of pioglitazone on either diastolic function or myocardial insulin sensitivity between drug-naïve and metformin-treated subjects, but the number of subjects in each group was small. Other than having diabetes, all subjects were in good general health as determined by medical history, physical examination, screening blood tests, urinalysis, and a normal electrocardiogram. Body weight was stable (± 3 lb) for at least 3 months before study entry. All

subjects were normally active, and none participated in an excessively heavy exercise program. Other than metformin and/ or sulfonylurea, no subject was taking any medication known to affect glucose metabolism. Ten subjects were taking a statin, and 10 were taking an antihypertensive medication (ACE inhibitor n = 7, angiotensin receptor blocker n = 3, calcium channel blocker n = 1).

Twelve healthy subjects with normal glucose tolerance (NGT) (age 47.7 \pm 10.5 years, BMI 28.4 \pm 0.4 kg/m², HbA_{1c} $5.5 \pm 0.4\%$; eight males, four females, nine Mexican American, three Caucasian) served as the control group. The entry criteria for the control group were similar to those for the T2D group. The protocol was approved by institutional review board of the University of Texas Health Science Center (San Antonio, TX), and written informed consent was obtained from all subjects.

Table 1-Metabolic and cardiac MRI values obtained in control subjects with NGT and subjects with T2D before and after pioglitazone treatment

	NGT control	T2D baseline	T2DM after pioglitazone
Sex, n			
Male	9	10	10
Female	3	2	2
Age (years)	47.7 ± 10.5	50.7 ± 9.1	51.3 ± 9.1
BMI (kg/m ²)	28.4 ± 4.4	30.8 ± 4.3	31.3 ± 4.2
Body fat (%)	29.3 ± 8.6	31.9 ± 5.7	33.4 ± 6.1
HbA _{1c} (%)	5.5 ± 0.4	6.7 ± 1.3***	5.6 ± 0.8‡
Fasting plasma glucose (mg/dL)	93 ± 6	149 ± 48***	112 ± 23†
Fasting FFAs (mmol/L)	0.32 ± 0.1	$0.52 \pm 0.17***$	$0.30 \pm 0.14 \ddagger$
HDL cholesterol (mg/dL)	55.7 ± 9.8	38.8 ± 11.9***	41.5 ± 9.7†
Triacylglycerol (mg/dL)	128 ± 94	265 ± 155***	153 ± 74‡
Matsuda index of insulin sensitivity	8.7 ± 4.8	2.8 ± 1.9***	5.8 ± 3.4‡
Glucose infusion rate (mg/kg·min)	7.5 ± 2.8	3.4 ± 1.3***	5.8 ± 2.1‡
MGU (μmol/min · g)	0.38 ± 0.14	$0.24 \pm 0.14*$	$0.42 \pm 0.13 \ddagger$
Myocardial perfusion (mL/min · g)	0.83 ± 0.20	0.95 ± 0.16	$1.10 \pm 0.25 \dagger$
Systolic function			
Resting heart rate (beats/min)	63.3 ± 6.8	$78.1 \pm 10.5**$	71.3 ± 11.3
Cardiac index (L/min · m²)	2.85 ± 0.32	2.90 ± 0.70	2.91 ± 0.74
EF (%)	64.2 ± 4.7	60.7 ± 6.3	65.6 ± 6.9†
Stroke volume/BSA (mL/m²)	42.7 ± 5.0	37.7 ± 7.3*	41.7 ± 8.5†
Peak LV ejection rate/BSA	225 1 25	224 . 52	255 + 54
$(mL/s \cdot m^2)$	226 ± 36	224 ± 52	255 ± 54
Myocardial mass/BSA (g/m²)	60.2 ± 9.4	64.1 ± 8.5	61.4 ± 8.6
Diastolic function			
Transmitral E/A flow ratio	1.48 ± 0.37	1.04 ± 0.28**	1.25 ± 0.38‡
ESV/BSA (mL/m²)	26.4 ± 9.7	24.3 ± 5.4	21.2 ± 5.3
EDV/BSA (mL/m²)	69.1 ± 10.3	61.9 ± 9.1*	62.9 ± 9.3
PLVFR/BSA (mL/s · m²)	196 ± 33	171 ± 52*	212 ± 54‡

Data are mean \pm SD. *P < 0.05; **P < 0.01; ***P < 0.005 determined by unpaired t test for T2D baseline vs. NGT control. $^\dagger P <$ 0.05 and $^\dagger P <$ 0.01 for T2D baseline vs. T2D after pioglitazone (determined by paired t test).

care.diabetesjournals.org Clarke and Associates 3

Study Design

At baseline, HbA_{1c} and fasting plasma glucose (FPG), insulin, and lipid concentrations were measured. At 8:00 A.M., after a 10-h overnight fast, a 2-h oral glucose tolerance test (OGTT) (75 g) was performed, with measurements of plasma glucose, insulin, and C-peptide every 15 min. DEXA (Hologic, Waltham, MA) was performed to determine total body fat and lean mass.

Cardiac MRI Studies

Within 1 week after the OGTT, subjects returned for a cardiac MRI study to assess cardiac morphology and function (30). Within 3–7 days after the MRI study, subjects returned for a euglycemic insulin clamp study to measure whole-body (primarily reflects skeletal muscle) insulin sensitivity (31).

MRI was performed on a 3.0-T system (TIM Trio; Siemens Healthcare, Malvern, PA) with a six-channel anterior phasedarray torso coil and corresponding posterior coil elements (12 channels total). Axial and sagittal localizer views; standard cardiac two-, three-, and four-chamber views; and 7-mm-thick slices were obtained by using gradient echo sequence $(2.2 \times 1.3 \text{-mm}^2 \text{ pixel area})$. Cine imaging with retrospective gating was used with a balanced steady-state free precession sequence (repetition time/echo time 2.44/1.22 ms). Acquisitions consisted of 25–30 cardiac phases (matrix 224 imes288, field of view [FOV] 336 \times 430 mm², 1.5×1.5 -mm² pixel area), which varied slightly by subject body size and heart rate. Contiguous short-axis slices were acquired during repetitive breath-holds at end expiration. Mitral inflow images were obtained by using phase-contrast gradient-echo sequence with throughplane velocity encoding (V_{enc} = 100 cm/s) at the mitral valve (flip angle 10°, repetition time/echo time 5.8/3.6 ms). Phase-contrast gradient-echo slice thickness was 8 mm (typical FOV 228 imes430 mm², matrix 192 imes 102), producing $2.89 \times 2.89 \times 8.0$ -mm 3 pixel volumes (30).

Euglycemic Insulin Clamp and PET

Cardiac PET study was performed in subjects with T2D while concurrently undergoing the insulin clamp. Subjects reported to the Research Imaging Institute at 7:00 A.M. after a 10-h overnight fast. PET scans were performed in two-dimensional imaging mode by using an ECAT 931-08/12

PET scanner (Control Technology Inc., Knoxville, TN) with a 10.5-cm axial FOV and resolution of $8.4\times8.3\times6.6~\text{mm}^3$ full width at half maximum. After optimization of subject position, a 20-min transmission scan was performed after exposure of a retractable ^{68}Ge ring source to correct emission data for tissue attenuation of γ -photons. Next, [$^{15}\text{O}]\text{H}_2\text{O}$ (10.5 MBq/kg) was administered through the catheter over 20 s, and the PET scan (10 mL/min) was performed to measure myocardial blood flow (MBF) as previously described (31).

Before the euglycemic insulin clamp (32), a catheter (for blood withdrawal) was placed in a vein on the dorsum of the hand, which rested in a box heated to 60°C (30). A second catheter for infusion of test substances was inserted into an antecubital vein. During the 30 min before the start of the insulin clamp, three baseline blood samples were drawn at 15-min intervals to allow sufficient time for decay of ¹⁵O radioactivity. At time 0, a primed continuous insulin (40 mU \cdot m⁻² \cdot min⁻¹) infusion was started for 150 min, and blood samples were drawn at 5-30-min intervals. No glucose was infused until plasma glucose concentration declined to 100 mg/dL, at which level it was maintained by variable infusion of 20% glucose. At 90 min after the start of insulin, [150]H20 was infused over 20 seconds for repeat measurement of MBF. At 105 min, [18F]fluoro-2-deoxy-D-glucose ([18F]FDG) (185 MBq) was infused, and a dynamic PET scan was performed for measurement of myocardial glucose uptake (MGU) as previously described (33). All radioactive tracers were produced at the onsite cyclotron radiochemistry facility.

Pioglitazone Treatment

After completion of the above studies, subjects with T2D were started on pioglitazone 15 mg/day. After 2 weeks, the dose was increased to 30 mg/day for 2 weeks. At week 4, the dose was increased to 45 mg/day and continued for an additional 20 weeks (total treatment period 24 weeks). Subjects received dietary counseling before initiation of pioglitazone therapy and were asked to consume a standard weight-maintaining American Diabetes Association diet throughout. Patients returned for follow-up every 2–4 weeks. After 24 weeks, baseline studies were repeated.

Data Analysis

MRI data were analyzed blindly by one of the investigators (G.D.C.), who used a commercial postprocessing package (cmr⁴²; Circle Cardiovascular Imaging, Calgary, AB, Canada). The cmr⁴² function module performed global and regional LV function analyses on slices acquired in short-axis orientation. LV volumes and myocardial mass were calculated with trabeculae and papillary muscles included. The cmr⁴² flow module computed velocity, flow, regurgitant volumes, and cardiac output. Ejection fraction (EF) was computed by using end-diastolic volume (EDV) and end-systolic volume (ESV). For determining differences between the control and pretreatment T2D groups, dimensional parameters were normalized to body surface area (BSA) by using the Mosteller equation (34). Ejection and filling functions were assessed from the respective maximal and average downslope and upslope of volume time curves to determine the peak LV ejection rate and peak LV filling rate (PLVFR).

PET sinograms were corrected for tissue attenuation and reconstructed through standard reconstruction algorithms. Image manipulation and data handling were performed by using MATLAB software (MathWorks, Natick, MA) (35). The input function for [18F]FDG was derived from continuous monitoring of arterialized blood radioactivity. Whole blood was converted into plasma input by using a limited number of discrete plasma samples. Delay correction was performed. Arterial input for [150]H20 was obtained from the left atrium timeactivity curve. The glucose infusion rate during the last 30 min of insulin clamp was stable in all studies and was averaged to obtain a measure of total-body insulinmediated glucose disposal.

Statistical Analysis

Data are expressed as mean \pm SD or as percentages. Statistical analyses were performed by using R version 3.2.1 statistical software with RStudio integrated development environment version 0.99.467. Unpaired two-sided Student t test was used to evaluate the null hypothesis of no difference between the baseline (prepioglitazone) T2D and control groups. Paired two-sided Student t test was used to evaluate the null hypothesis of no difference between the baseline (prepioglitazone) and posttreatment

pioglitazone studies in T2D. P < 0.05 was deemed significant.

RESULTS

The control and T2D groups were similar in age, BMI, body fat, and sex (Table 1). HbA_{1c}, FPG, and fasting plasma free fatty acids (FFAs) were lower in the control versus T2D group at baseline (P < 0.005). The Matsuda index of insulin sensitivity was greater in the control versus T2D group at baseline (P < 0.001) (Table 1).

EDV and ESV, normalized to BSA, were greater in control subjects (69.1 ± 10.3 mL/m² and 26.4 \pm 9.7 mL/m², respectively; both P < 0.05) versus subjects with T2D (61.9 \pm 9.1 mL/m² and 24.3 \pm 5.4 mL/m², respectively). Early diastolic relaxation/atrial contraction (E/A) ratio was significantly higher (P < 0.01) in the control (1.48 \pm 0.37) versus T2D (1.04 \pm 0.28) group. PLVFR/BSA was significantly greater (P < 0.05) in control subjects $(196 \pm 33 \text{ mL/s} \cdot \text{m}^2)$ versus subjects with T2D (171 \pm 52 mL/s \cdot m²).

In the subjects with T2D, pioglitazone reduced HbA_{1c} (6.7 \pm 1.3 to 5.6 \pm 0.8%; P< 0.01), FPG (149 \pm 48 to 112 \pm 23 mg/dL; P < 0.05), and fasting plasma FFAs $(0.52 \pm 0.17 \text{ to } 0.30 \pm 0.14 \text{ mmol/L};$ P < 0.01) (Table 1). Insulin-stimulated whole-body (primarily reflects skeletal muscle) glucose uptake increased from $3.4 \pm 1.3 \text{ to } 5.8 \pm 2.1 \text{ mg/kg} \cdot \text{min } (P < 1.3 \text{ to } 1.3 \text{ to }$ 0.01), and the Matsuda index of insulin sensitivity during OGTT increased from 2.8 ± 1.9 to 5.8 ± 3.4 (P < 0.01). After pioglitazone treatment, MGU (measured with [18F]FDG PET scan), increased from 0.24 \pm 0.14 to 0.42 \pm 0.13 $\mu mol/min \cdot g$ (P < 0.01), and MBF (measured with [150]H₂O PET scan) increased from 0.95 \pm 0.16 to 1.10 \pm 0.25 mL/min \cdot g tissue (P < 0.05).

After pioglitazone treatment, both systolic (124 \pm 12 to 117 \pm 10 mmHg) and diastolic (80 \pm 9 to 73 \pm 9 mmHg) blood pressure were reduced (P < 0.05). Percent body fat and BMI increased slightly (P not significant) after pioglitazone (Table 1). Edema was observed in 2 of the 12 subjects treated with pioglitazone and was considered to be mild in both.

Also after pioglitazone treatment, parameters of systolic function (determined by cardiac MRI) improved. Stroke volume increased from 37.7 \pm 7.3 to 41.7 \pm 8.5 mL/m² (P < 0.05), and EF increased from 60.7 \pm 5.1 to 65.6.0 \pm 6.9% (P < 0.05). Resting heart rate decreased

modestly (P not significant). Diastolic dysfunction improved after pioglitazone. Transmitral E/A ratio (1.04 \pm 0.28 to 1.25 \pm 0.38; P < 0.01) and PLVFR $(171 \pm 52 \text{ to } 212 \pm 54 \text{ mL/s} \cdot \text{m}^2; P <$ 0.01) both increased markedly. No significant differences in LV volumes or myocardial mass were found (Table 1).

Change in HbA_{1c} correlated inversely with insulin-stimulated glucose disposal during insulin clamp (r = -0.47; P < 0.05) and with the change in MGU (r = -0.64; P = 0.02) after pioglitazone treatment. The decrement in plasma FFAs also correlated with the change in MGU (r = -0.67; P = 0.02) (Supplementary Fig. 1).

Insulin-stimulated whole-body (muscle) glucose uptake during insulin clamp correlated positively and strongly with MGU (r = 0.50; P = 0.01) (Fig. 1A) and both measures of diastolic function (transmitral E/A ratio: r = 0.52; P = 0.01; PLVFR: r = 0.55; P = 0.005) (Fig. 1A and B). The Matsuda index of insulin sensitivity correlated positively with MGU (r = 0.51; P = 0.01).

The increase in E/A ratio after pioglitazone treatment was strongly correlated with the reduction in HbA_{1c} (r = -0.74; P < 0.01) (Fig. 2A), increase in MGU (r =0.51; P = 0.03) (Fig. 2B), and increase in insulin-stimulated whole-body glucose uptake during insulin clamp (r = 0.58; P = 0.047) (Fig. 2C). The change in PLVFR after pioglitazone treatment correlated with change in HbA_{1c} (r = -0.56; P <0.05) and tended to correlate with the change in MGU (r = 0.51; P = 0.09) and insulin-stimulated whole-body glucose uptake (r = 0.46; P = 0.13) during insulin clamp (Fig. 3A-C). The change in E/A ratio correlated strongly with PLVFR after pioglitazone treatment (r = 0.69; P < 0.001).

CONCLUSIONS

The current study provides three novel findings. To our knowledge, this study is

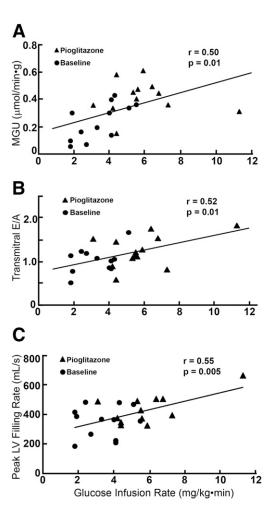


Figure 1—Correlation between insulin-stimulated whole-body glucose disposal (glucose infusion rate) and MGU (A), transmitral E/A ratio (B), and PLVFR (C) in subjects with T2D before (●) and after (A) pioglitazone treatment.

care.diabetesjournals.org Clarke and Associates 5

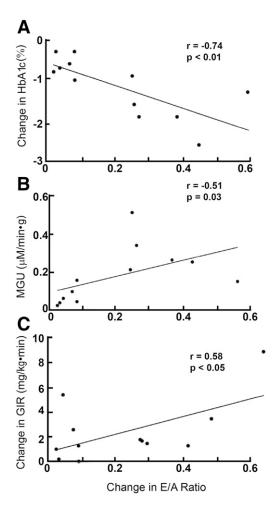


Figure 2—Correlation between the change in transmitral E/A ratio and change in HbA_{1c} (A), MGU (B), and glucose infusion rate (GIR) (C) during insulin clamp after pioglitazone treatment.

the first to 1) simultaneously quantitate myocardial and skeletal muscle insulin resistance in the same subject and to document a strong correlation between the two; 2) demonstrate that pioglitazone or any thiazolidinedione improves both diastolic and systolic function in patients with T2D without clinical CV disease; and 3) demonstrate that the improvements in diastolic and systolic function are closely related to improved myocardial (and skeletal muscle) insulin sensitivity after pioglitazone treatment. Consistent with previous results (8), the current study demonstrates that pioglitazone does not exert any negative effects on cardiac function in patients with T2D without clinically manifest CV disease and that diastolic dysfunction is present early in the natural history of these patients.

Insulin resistance in skeletal muscle is a characteristic feature of T2D (7). Myocardial insulin resistance also has been demonstrated

in individuals with T2D with and without coronary artery disease (8,9). Diastolic dysfunction and increased LV diastolic stiffness are commonly observed in T2D. These abnormalities are observed early in the natural history of T2D and have been related to underlying myocardial insulin resistance (4,5,19). FFAs are the major metabolic fuel for the heart; however, under conditions of ischemia, the heart switches to glucose utilization because of the higher ATP yield per amount of oxygen consumed (5). The diabetic heart has been postulated to lose its metabolic flexibility because of myocardial insulin resistance (18,19), and this is associated with myocardial lipid accumulation, inflammation, increased collagen formation, myocardial stiffness, and a noncompliant LV (4,5,8,19). Thiazolidinediones are potent insulin sensitizers in skeletal muscle (7,36), and both rosiglitazone (37) and pioglitazone (8) augment insulinmediated MGU in patients with T2D. In the rosiglitazone study (37), no measures of myocardial function were performed, and in the pioglitazone study (8), no correlation between improved myocardial insulin resistance and cardiac function was observed. In diabetic rodents treated with pioglitazone, a reduction in myocardial collagen content was demonstrated in association with improved LV function (38). Collectively, these results suggest that pioglitazone improves LV diastolic dysfunction, but whether this improvement is related to the increase in myocardial insulin sensitivity/MGU is unclear. On the other hand, pioglitazone therapy is associated with fluid retention (39,40) and can lead to heart failure, especially in patients with T2D and underlying diastolic dysfunction (20), raising concern about the use of thiazolidinediones in T2D. However, salt and water retention with peroxisome proliferator-activated receptor-y agonists is related to enhanced renal sodium reabsorption and, as demonstrated by the current results, not to a negative effect on the myocardium. To the contrary, pioglitazone improved all parameters of LV diastolic dysfunction, including the transmitral E/A ratio, PLVFR, and EDV (Table 1). Key parameters of systolic function also were enhanced with pioglitazone. Thus, in patients with T2D with diastolic dysfunction but without clinically evident CV disease, pioglitazone improved parameters of diastolic as well as systolic LV function. These findings are consistent with an echocardiographic study in patients with T2D (41). Of note, consistent with previous results (24,42), pioglitazone reduced both systolic and diastolic blood pressure while reducing heart rate. Thus, despite pioglitazone's renal sodium retentive effect, blood pressure dropped most likely secondary to vasodilation, leading to afterload reduction, and this could partly explain the beneficial effect of pioglitazone on both LV systolic and LV diastolic function.

Although the insulin sensitizing effect of thiazolidinediones on skeletal muscle is well established (7), the effect of thiazolidinediones on myocardial insulin sensitivity has been less well studied (8,33). In the current study, pioglitazone improved skeletal muscle and myocardial insulin sensitivity in subjects with T2D by 71% and 75%, respectively, and a strong correlation between the these increases was observed (P = 0.50 and P < 0.01,

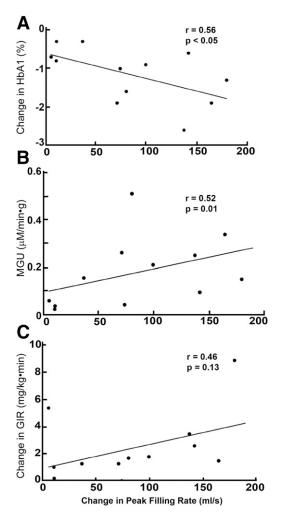


Figure 3—Correlation between the change in PLVFR and change in $HbA_{1c}(A)$, MGU (B), and glucose infusion rate (GIR) (C) during insulin clamp after pioglitazone treatment.

respectively) (Fig. 1). The myocardial insulin sensitizing effect of pioglitazone is consistent with that reported previously (8,37), but these studies did not measure skeletal muscle insulin sensitivity. The current results indicate that both skeletal and cardiac muscle insulin resistance is characteristic of T2D. Of note, our results demonstrate for the first time to our knowledge a strong correlation between improved myocardial (and skeletal muscle) insulin sensitivity and improved diastolic dysfunction, as manifested by the increased E/A ratio (Fig. 2) and PLVFR (Fig. 3). The study by van der Meer et al. (8) also showed increases in myocardial insulin sensitivity and LV diastolic function in pioglitazone-treated patients but did not find a significant correlation between these two variables. These authors' failure to detect a correlation stands in contrast to the current results and may be explained by the higher daily dose of pioglitazone (45 vs. 30 mg) used in the current study or by the difference in study design and/or concomitant antidiabetic therapy (10-week washout followed by a switch to glimepiride monotherapy).

Both lipotoxicity and glucotoxicity contribute to skeletal muscle insulin resistance in T2D (7). The current results suggest that these same pathogenic factors also play a role in diabetic myocardial insulin resistance. Both the decrement in HbA_{1c} (r = -0.64; P = 0.02) and the decrement in plasma FFAs (r = -0.67; P = 0.02) correlated with enhanced insulinstimulated myocardial glucose uptake after pioglitazone treatment (Supplementary Fig. 1). Similarly, improvements in E/A ratio and PLVFR were related to the reduction in HbA_{1c} (r = -0.74 and -0.56, respectively; both P < 0.05) (Figs. 2 and 3). In the current study, we observed a 21% increase in insulin-stimulated MBF after pioglitazone treatment, and thiazolidinediones have been shown to augment insulin signaling in skeletal muscle (43). Thus, multiple factors (improved glycemic control, reduced plasma FFAs, increased MBF, increased insulin signaling) could have contributed to the improvement in myocardial insulin sensitivity, whereas both amelioration of myocardial insulin resistance and decrease in blood pressure likely contributed to enhanced LV diastolic and systolic function.

The study has several limitations. First, the number of subjects was relatively small. Second, we specifically enrolled patients with T2D who did not have clinically manifest cardiac disease. Whether similar results would be observed in T2D with longer duration of diabetes and clinical evidence of cardiac dysfunction remains to be examined. Third, radiolabeled glucose was not used; therefore, insulinstimulated whole-body (skeletal muscle) glucose uptake may have been underestimated in the subjects with T2D during baseline insulin clamp because of incomplete suppression of endogenous glucose production. However, even if endogenous glucose production continued at 0.5-1.0 mg/kg · min (the latter value is unlikely), pioglitazone treatment still would have significantly increased skeletal muscle insulin sensitivity.

In summary, the results demonstrate that 6 months of pioglitazone treatment in patients with T2D without clinically evident CV disease ameliorates myocardial insulin resistance, augments MBF, improves both LV diastolic and LV systolic function, and reduces blood pressure while decreasing heart rate. These observations indicate that 1) LV diastolic dysfunction in T2D is closely correlated with myocardial insulin resistance and that both are improved by pioglitazone and 2) pioglitazone exerts no negative effects on myocardial function and can be used safely in patients with T2D without clinically evident CV disease.

Funding. This work was supported by National Institute of Diabetes and Digestive and Kidney Diseases grant R01-DK-2409333. The salary of R.A.D. is supported in part by the South Texas Veterans Health Care System.

Duality of Interest. This work was supported by a grant from Takeda Pharmaceuticals North America. E.C. is on the speaker's bureaus of Janssen, Eli Lilly, Boehringer Ingelheim, AstraZeneca, and Sanofi and receives research support from AstraZeneca and Janssen. R.J.C. is an advisory board member for Pfizer, Boehringer Ingelheim, AstraZeneca, Boston Scientific, Merck Sharp & Dohme, and

care.diabetesjournals.org Clarke and Associates 7

Takeda Pharmaceuticals. A.G. is a consultant for Eli Lilly, Menarini, and Genentech. R.A.D. is on the advisory boards of AstraZeneca, Novo Nordisk, Janssen, Intarcia, Elcelyx, and Boehringer Ingelheim; receives research support from Boehringer Ingelheim, Takeda Pharmaceuticals, Janssen, and AstraZeneca; and is on the speaker's bureaus of Novo Nordisk and AstraZeneca. No other potential conflicts of interest relevant to this article were reported.

Author Contributions. G.D.C., C.S.-H., M.M.-W., S.M., A.M., E.C., R.J.C., P.I., A.G., M.A.-G., and R.A.D. performed the study and reviewed the manuscript. G.D.C. and R.A.D. designed the study and wrote the first draft of the manuscript. R.A.D. is the guarantor of this work and, as such, had full access to all the data in the study and takes responsibility for the integrity of the data and the accuracy of the data analysis.

References

- 1. Beckman JA, Creager MA, Libby P. Diabetes and atherosclerosis: epidemiology, pathophysiology, and management. JAMA 2002;287:2570–2581
- 2. Haffner SM, Lehto S, Rönnemaa T, Pyörälä K, Laakso M. Mortality from coronary heart disease in subjects with type 2 diabetes and in nondiabetic subjects with and without prior myocardial infarction. N Engl J Med 1998;339:229–234
- 3. Bhuiyan T, Maurer MS. Heart failure with preserved ejection fraction: persistent diagnosis, therapeutic enigma. Curr Cardiovasc Risk Rep 2011;5: 440–449
- 4. Diamant M, Lamb HJ, Groeneveld Y, et al. Diastolic dysfunction is associated with altered myocardial metabolism in asymptomatic normotensive patients with well-controlled type 2 diabetes mellitus. J Am Coll Cardiol 2003;42:328–335
- 5. Boudina S, Abel ED. Diabetic cardiomyopathy revisited. Circulation 2007;115:3213–3223
- 6. Gilbert RE, Krum H. Heart failure in diabetes: effects of anti-hyperglycaemic drug therapy. Lancet 2015;385:2107–2117
- 7. DeFronzo RA. Banting Lecture. From the triumvirate to the ominous octet: a new paradigm for the treatment of type 2 diabetes mellitus. Diabetes 2009;58:773–795
- van der Meer RW, Rijzewijk LJ, de Jong HW, et al. Pioglitazone improves cardiac function and alters myocardial substrate metabolism without affecting cardiac triglyceride accumulation and high-energy phosphate metabolism in patients with well-controlled type 2 diabetes mellitus. Circulation 2009:119:2069–2077
- 9. lozzo P, Chareonthaitawee P, Dutka D, Betteridge DJ, Ferrannini E, Camici PG. Independent association of type 2 diabetes and coronary artery disease with myocardial resonance spectroscopy study. Circulation 2002;116:1170–1175 10. Gastaldelli A, Cusi K, Pettiti M, et al. Relationship between hepatic/visceral fat and hepatic insulin resistance in nondiabetic and type 2 diabetic subjects. Gastroenterology 2007;133:496–506
- 11. Groop LC, Bonadonna RC, DelPrato S, et al. Glucose and free fatty acid metabolism in non-insulin-dependent diabetes mellitus. Evidence for multiple sites of insulin resistance. J Clin Invest 1989;84:205–213
- 12. Bajaj M, Baig R, Suraamornkul S, et al. Effects of pioglitazone on intramyocellular fat metabolism in patients with type 2 diabetes mellitus. J Clin Endocrinol Metab 2010;95:1916–1923

13. Yki-Järvinen H. Thiazolidinediones. N Engl J Med 2004:351:1106–1118

- 14. Miyazaki Y, Mahankali A, Matsuda M, et al. Improved glycemic control and enhanced insulin sensitivity in type 2 diabetic subjects treated with pioglitazone. Diabetes Care 2001;24:710–719
- 15. Bajaj M, Suraamornkul S, Hardies LJ, Glass L, Musi N, DeFronzo RA. Effects of peroxisome proliferator-activated receptor (PPAR)-alpha and PPAR-gamma agonists on glucose and lipid metabolism in patients with type 2 diabetes mellitus. Diabetologia 2007;50:1723–1731
- 16. Schernthaner G, Currie CJ, Schernthaner GH. Do we still need pioglitazone for the treatment of type 2 diabetes? A risk-benefit critique in 2013. Diabetes Care 2013;36(Suppl. 2):S155–S161
- 17. DeFronzo RA. Insulin resistance, lipotoxicity, type 2 diabetes and atherosclerosis: the missing links. The Claude Bernard Lecture 2009. Diabetologia 2010;53:1270–1287
- 18. Nolan CJ, Ruderman NB, Kahn SE, Pedersen O, Prentki M. Insulin resistance as a physiological defense against metabolic stress: implications for the management of subsets of type 2 diabetes. Diabetes 2015;64:673–686
- 19. Taegtmeyer H, Golfman L, Sharma S, Razeghi P, van Arsdall M. Linking gene expression to function: metabolic flexibility in the normal and diseased heart. Ann N Y Acad Sci 2004;1015:202–213 20. Dormandy JA, Charbonnel B, Eckland DJ, et al.; PROactive Investigators. Secondary prevention of macrovascular events in patients with type 2 diabetes in the PROactive study (PROspective pioglitAzone Clinical Trial In macroVascular Events): a randomised controlled trial. Lancet 2005;366: 1279–1289
- 21. Dormandy JA, Betteridge DJ, Schernthaner G, Pirags V, Norgren L; PROactive Investigators. Impact of peripheral arterial disease in patients with diabetes—results from PROactive (PROactive 11). Atherosclerosis 2009;202:272–281
- 22. UK Prospective Diabetes Study (UKPDS) Group. Intensive blood-glucose control with sulphonylureas or insulin compared with conventional treatment and risk of complications in patients with type 2 diabetes (UKPDS 33). Lancet 1998;352:837–853
- 23. Collins R, Armitage J, Parish S, Sleigh P, Peto R; Heart Protection Study Collaborative Group. MRC/BHF Heart Protection Study of cholesterollowering with simvastatin in 5963 people with diabetes: a randomised placebo-controlled trial. Lancet 2003:361:2005–2016
- 24. McMurray JJ, Gerstein HC, Holman RR, Pfeffer MA. Heart failure: a cardiovascular outcome in diabetes that can no longer be ignored. Lancet Diabetes Endocrinol 2014;2:843–851
- 25. Kernan WN, Viscoli CM, Furie KL, et al.; IRIS Trial Investigators. Pioglitazone after ischemic stroke or transient ischemic attack. N Engl J Med 2016;374:1321–1331
- 26. Nesto RW, Bell D, Bonow RO, et al. Thiazolidinedione use, fluid retention, and congestive heart failure: a consensus statement from the American Heart Association and American Diabetes Association. Diabetes Care 2004;27:256–263 27. Home P. Safety of PPAR agonists. Diabetes Care 2011;34(Suppl. 2):S215–S219
- 28. Garber AJ, Abrahamson MJ, Barzilay JI, et al. Consensus statement by the American Association of Clinical Endocrinologists and American College of Endocrinology on the comprehensive type 2

diabetes management algorithm – 2017 executive summary. Endocr Pract 2017;23:207–238

- 29. American Diabetes Association. Standards of medical care in 2017. Diabetes Care 2017;40 (Suppl. 1):S1–S132
- 30. Hartiala JJ, Mostbeck GH, Foster E, et al. Velocity-encoded cine MRI in the evaluation of left ventricular diastolic function: measurement of mitral valve and pulmonary vein flow velocities and flow volume across the mitral valve. Am Heart J 1993:125:1054–1066
- 31. Iozzo P, Chareonthaitawee P, Dutka D, Betteridge DJ, Ferrannini E, Camici PG. Independent association of type 2 diabetes and coronary artery disease with myocardial insulin resistance. Diabetes 2002;51:3020–3024
- 32. DeFronzo RA, Tobin JD, Andres R. Glucose clamp technique: a method for quantifying insulin secretion and resistance. Am J Physiol 1979;237: E214–E223
- 33. Lautamäki R, Airaksinen KE, Seppänen M, et al. Rosiglitazone improves myocardial glucose uptake in patients with type 2 diabetes and coronary artery disease: a 16-week randomized, double-blind, placebo-controlled study. Diabetes 2005;54:2787–2794
- 34. Verbraecken J, Van de Heyning P, De Backer W, Van Gaal L. Body surface area in normal-weight, overweight, and obese adults. A comparison study. Metabolism 2006;55:515–524
- 35. Hermansen F, Ashburner J, Spinks TJ, Kooner JS, Camici PG, Lammertsma AA. Generation of myocardial factor images directly from the dynamic oxygen-15-water scan without use of an oxygen-15-carbon monoxide blood-pool scan. J Nucl Med 1998;39:1696–1702
- 36. Eldor R, DeFronzo RA, Abdul-Ghani M. In vivo actions of peroxisome proliferator-activated receptors: glycemic control, insulin sensitivity, and insulin secretion. Diabetes Care 2013;36(Suppl. 2): \$162–\$174
- 37. Hällsten K, Virtanen KA, Lönnqvist F, et al. Enhancement of insulin-stimulated myocardial glucose uptake in patients with type 2 diabetes treated with rosiglitazone. Diabet Med 2004;21:1280–1287 38. Tsuji T, Mizushige K, Noma T, et al. Pioglitazone improves left ventricular diastolic function and decreases collagen accumulation in prediabetic stage of a type II diabetic rat. J Cardiovasc Pharmacol 2001;38:868–874
- 39. Pavlov TS, Imig JD, Staruschenko A. Regulation of EnaC-mediated sodium reabsorption by peroxisome proliferator-activated receptors. PPAR Res 2010;2010:703735
- 40. Guan Y, Hao C, Cha DR, et al. Thiazolidinediones expand body fluid volume through PPARgamma stimulation of ENaC-mediated renal salt absorption. Nat Med 2005;11:861–866
- 41. Ordu S, Ozhan H, Alemdar R, et al. Pioglitazone improves ventricular diastolic function in patients with diabetes mellitus: a tissue Doppler study. Acta Cardiol 2010;65:401–406
- 42. Derosa G, Fogari E, Cicero AF, et al. Blood pressure control and inflammatory markers in type 2 diabetic patients treated with pioglitazone or rosiglitazone and metformin. Hypertens Res 2007;30:387–394
- 43. Miyazaki Y, He H, Mandarino LJ, DeFronzo RA. Rosiglitazone improves downstream insulin receptor signaling in type 2 diabetic patients. Diabetes 2003;52:1943–1950